

Report 3

Effects of a Late-Summer Grassland Fire on Diffuse Knapweed Density

3. Effects of a Late-Summer Grassland Fire on Diffuse Knapweed Density

3.1 Introduction

Diffuse knapweed is listed as a noxious weed under Colorado State law (CRS 1996) and as a result must be controlled by property owners. Prescribed burns have been proposed as a possible component of a comprehensive program to manage the ecological resources of the Site. However, diffuse knapweed is a species adapted to disturbance regimes, and concern has been expressed that a fire disturbance might actually promote the spread of the species (Sheley et al. 1998). A natural fire in 1996 provided an opportunity to evaluate the effects of a burn on an area of diffuse knapweed infestation.

On September 2, 1996, lightning caused a wildfire that swept across approximately 105 acres of grassland in the Buffer Zone south of the Industrial Area at Rocky Flats Environmental Technology Site (Site). Observations the next day suggested that the fire was a fast-moving, cool fire. While most of the litter and much of the live biomass had been removed, diffuse knapweed (*Centaurea diffusa*) was not consumed and remained standing. The adult plants had only been scorched at the base, leaving current-year stalks still standing, and rosettes that were present before the fire had only their leaf tips scorched. As a result, it was possible to determine pre-burn stem densities.

This study was designed to examine the effects of the late summer grassland fire on the stem densities of diffuse knapweed. The hypothesis being tested was:

H_0 = There would be no difference in the pre-burn and post-burn stem density of diffuse knapweed (post-burn being two years after the fire).

3.2 Methods

The study was designed using both unburned (control) and burned (treatment) plots where diffuse knapweed was present in visually similar amounts. Where possible, the control and treatment plots were selected adjacent to each other across the edge of the fire line. This was possible for only a portion of the diffuse knapweed control plot. The remainder of the control plot was located across a gravel road where the fire had burned up to the road edge. The plant community was the same, however. The soil type present at the study location is Flatirons very cobbly sandy loam (SCS 1980).

Ten replicate, square-shaped 1-m² quadrats were located randomly in each of the unburned and burned plots. Each quadrat location was staked with rebar at one corner, and quadrats were oriented using a compass so that the edges were aligned N-S and E-W. The staked corner position for each quadrat was recorded, so the quadrat could be

relocated accurately for future sampling. Stem densities were counted and recorded for each quadrat. Both adult plants (stalks) and rosettes were counted, and their numbers were summed for the total stem density per quadrat. All three values were recorded for each quadrat. Adult plants were defined as all plants that had bolted. Rosette counts included both seedling and rosette growth forms. Sampling was conducted in late summer 1996, 1997, and 1998.

Data were entered and quality checked prior to analysis. Data were summarized using the 10 quadrats sampled for each plot ($n = 10$). Pre- and post-burn stem-density analyses between the control and treatment plots were conducted using a Mann-Whitney U test ($P = 0.05$, two-tailed test; Fowler and Cohen, 1990). Analyses of between-year differences in stem densities within treatment types were conducted using Wilcoxon's test for matched pairs ($P = 0.05$, two-tailed test; Fowler and Cohen, 1990). Both tests are ranking tests that compare the medians of the samples. Statistical analyses used only the 1996 and 1998 data.

3.3 Results

The results for the unburned and burned sites are shown in Table 3-1 and Figures 3-1 and 3-2. Diffuse knapweed rosette densities increased significantly in both the unburned and burned plots from 1996 to 1998 ($P = 0.05$; Table 3-1 and Figure 3-1). Although rosette density was higher in the burned plot than in the unburned plot for each of the three years, the difference in rosette density was not statistically significant in 1996 or 1998 ($P = 0.05$; 1997 not analyzed; Table 3-1 and Figure 3-1). The number of adult plants also increased in both the unburned plot and burned plot from 1996 to 1998, with the burned plot having the higher density during each year (Table 3-1 and Figure 3-2). Although there was a statistically significant increase in the density of adult plants in the burned plot over this time, the difference between plots was not statistically significant in either 1996 or 1998 ($P = 0.05$; 1997 not analyzed; Table 3-1 and Figure 3-2).

The number of rosettes in the unburned and burned plots increased by factors of 7 and 10, respectively, over the two-year period (Table 3-1 and Figure 3-1). The number of adult plants increased by factors of 2.7 and 3.3 in the unburned and burned plots, respectively, for the same two years (Table 3-1 and Figure 3-2). From 1996 to 1997, the number of diffuse knapweed rosettes increased by factors of approximately 3 and 2, respectively, for the unburned and burned plots (Table 3-1 and Figure 3-1). At the same time, the number of adult plants doubled in the burned plots while remaining the same in the unburned plots (Table 3-1 and Figure 3-2). The number of adult plants nearly doubled again from 1997 to 1998 in the burned plots, while nearly tripling in the unburned plots during the same time period (Table 3-1 and Figure 3-2).

3.4 Discussion

The diffuse knapweed response in the unburned and burned plots were generally parallel, with both the rosette and adult plant life stages showing increased densities over time.

The fact that diffuse knapweed rosette densities increased significantly in both the unburned and burned plots from 1996 to 1998, combined with the lack of a statistically significant difference ($P = 0.05$) in the rosette densities between the unburned and burned plots in 1996 and again in 1998, suggests that the late-summer grassland fire had a negligible impact on the rosette density of diffuse knapweed. The data, however, do illustrate how rapidly diffuse knapweed can increase in an infested area, and that the potential for large increases in diffuse knapweed density are present with or without fire in otherwise undisturbed native plant communities..

The effect of fire on diffuse knapweed infestations is a concern of land managers, because one of the most important tools for prairie management and restoration is fire. Results elsewhere on diffuse knapweed and spotted knapweed have shown that fire does not control these species, because the fires have not been hot enough to affect seed germination (Sheley et al. 1998). Spotted knapweed infestations have been shown to actually increase after prescribed burns (Sheley et al. 1998). However, the present study revealed significant increases in knapweed density in both the burned and unburned plots, indicating that diffuse knapweed populations will increase in an infested area whether it is burned or not. Therefore, the concern that using fire as a component of a comprehensive land management program will result in knapweed infestations beyond what would occur naturally is not justified, based on the results of this investigation.

Examined from a different point of view, the use of prescribed burns in conjunction with herbicide applications could potentially improve the effectiveness of these applications in controlling diffuse knapweed. In Montana's Lolo National Forest, prescribed burns in combination with aerial herbicide applications have been successful in controlling spotted knapweed infestations on steep mountain grasslands (Henry 1998). Locally, the City of Boulder Mountain Parks is conducting trials using prescribed burns followed by herbicide applications to control diffuse knapweed (Armstrong 1998, pers. comm.). Results of their trials appear promising and could potentially be used on the Site, in addition to current weed control methods.

As new management techniques are developed, these best management techniques will be pursued to control diffuse knapweed more effectively on the Site.

3.5 References

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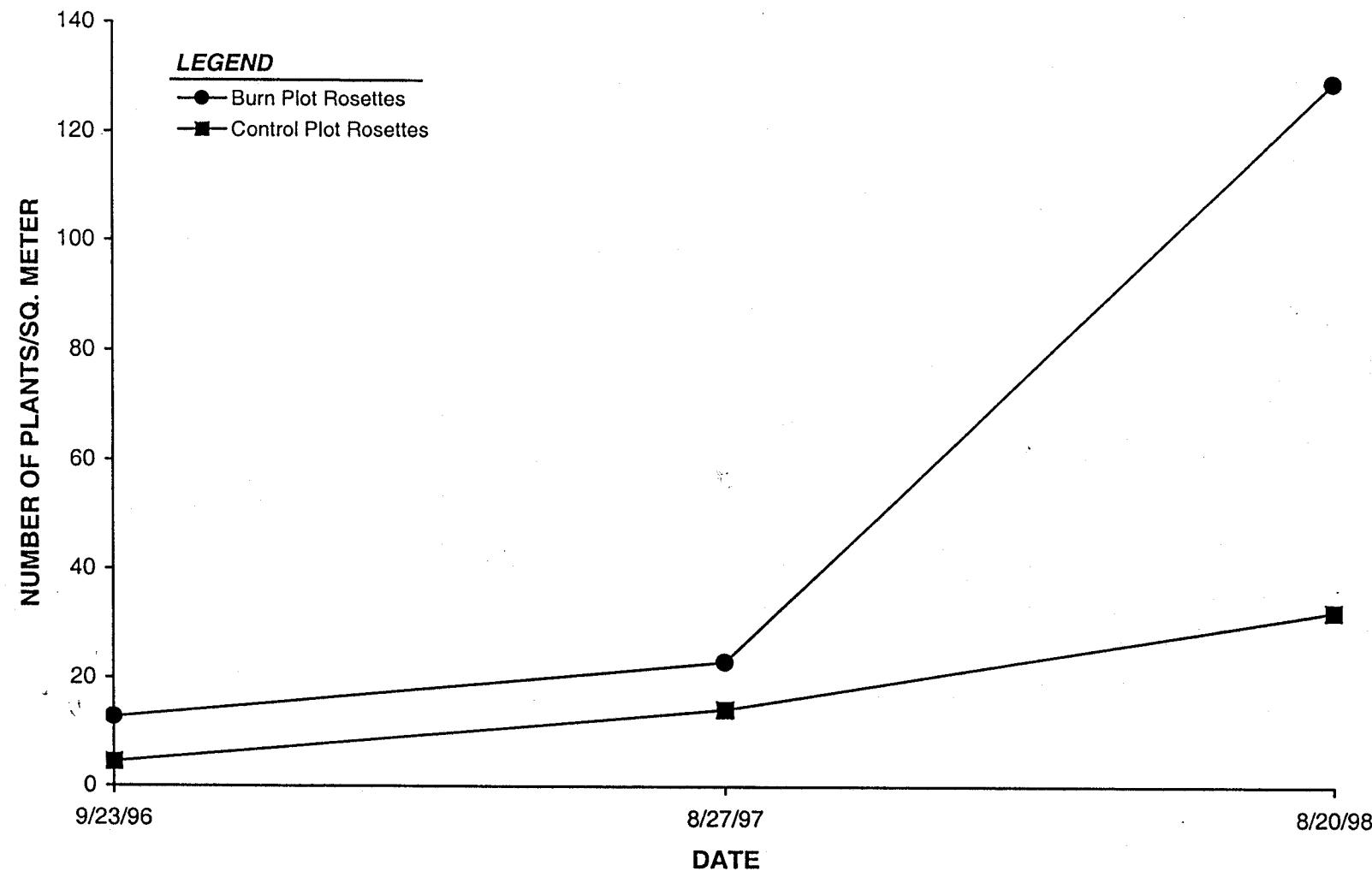


Figure 3-1. Diffuse knapweed rosette densities from 1996–1998.

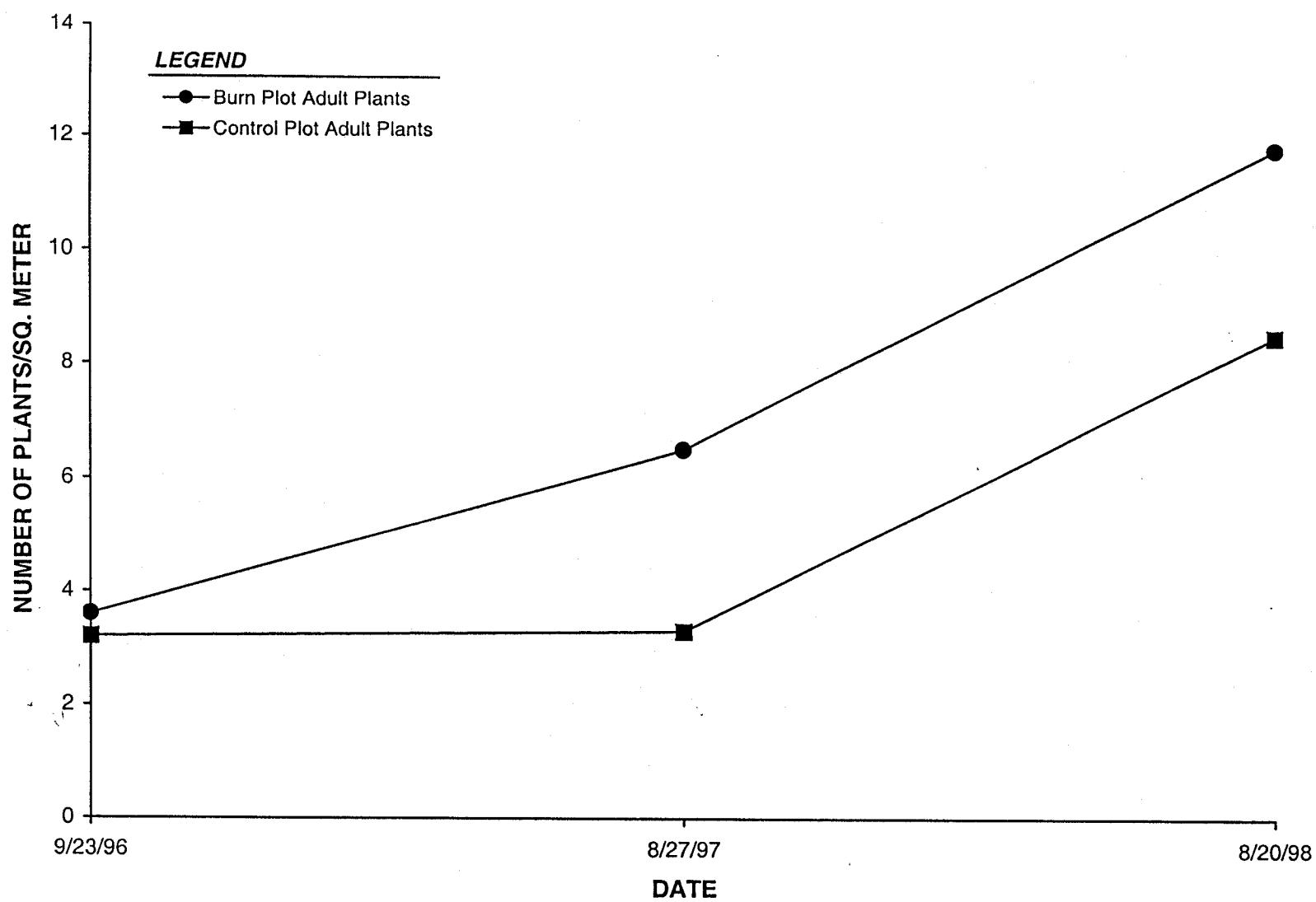


Figure 3-2. Diffuse knapweed adult plant densities from 1996–1998.

Table 3-1. 1996–1998 *Centaurea diffusa* densities in control and burn plots

Sampsite	Data	Plant Density (mean # of plants/sq. meter)		
		9/23/96	8/27/97	8/20/98
Burn plot	Rosettes	12.8	23	129.3
Control plot	Rosettes	4.5	14.3	32.4
Burn plot	Mature plants	3.6	6.5	11.8
Control plot	Mature plants	3.2	3.3	8.5
Burn plot	Total # plants	16.4	29.5	141.1
Control plot	Total # plants	7.7	17.6	40.9

Report 4

1998 Wetland Monitoring Summary

4. 1998 Wetland Monitoring Summary

4.1 Introduction

Monitoring is an integral part of determining whether the management objectives and goals for the high-value plant communities at Rocky Flats Environmental Technology Site (Site) are being achieved (IMP 1997; K-H 1997a,b). Consistent with this goal, long-term quantitative monitoring is necessary to determine whether changes are taking place in the plant communities that might go undetected through the use of broader-scale qualitative monitoring techniques.

Wetlands are an integral part of the overall Site ecosystem. Their numerous functions include providing habitat for many plants and animals, storing and releasing water, maintaining water quality, and controlling erosion (COE 1994). In 1994, the U.S. Army Corps of Engineers (COE) conducted a wetlands inventory that identified more than 1,000 wetlands on the Site, covering 191 acres (COE 1994). The Site vegetation map, updated in 1996 using a broader definition of wetlands areas, identified approximately 407 acres of wetlands on the Site. The broader definition used for the 1996 map allowed inclusion of wet meadow areas.

At present, the only quantitative baseline data available for wetlands on the Site is from a few scattered localities where sampling was conducted during the baseline inventory study in 1991 (DOE 1992). The COE inventory in 1994 delineated wetlands on the Site, but gathered only qualitative vegetation data (COE 1994). During 1997, as part of the high-value vegetation community monitoring, species richness was inventoried in three of the largest wetland areas on the Site (K-H 1997c), documenting 260 species of vascular plants in these wetlands (K-H 1998). In the 1998 study, 15 permanently marked transects were located in the wetlands that had been qualitatively surveyed in 1997, to provide quantitative long-term monitoring data for the wetland community (Figure 4-1). Monitoring was conducted using the same methods as were used for other quantitative vegetation monitoring on the Site, allowing compatibility with and comparability to other Site data. The purpose of this monitoring was to provide quantitative baseline information at permanent wetland locations that could be used to assess and document future changes in these communities.

The following questions were proposed for the 1998 monitoring, to provide baseline information on the species composition in the wetland communities on the Site:

1. What is the baseline species richness at these wetland locations?
2. What is the baseline foliar cover (total and individual species) at these wetland locations?

3. What is the baseline percent of total native foliar cover at these wetland locations?
4. What is the baseline woody plant densities at these wetland locations?
5. What is the baseline frequency of occurrence of individual species at these wetland locations?
6. How does the species richness and foliar cover information from the wetlands compare to that of the other plant communities on the Site?

4.2 Methods

4.2.1 Study Site Information

The areas selected for this study were two large wetlands located in Rock Creek and the Antelope Springs/Apple Orchard wetland in Woman Creek (Figure 4-1). These areas were chosen for two primary reasons: they are the largest wetlands on the Site, and a floristic inventory was conducted in each of these areas in 1997. The COE (1994) study that delineated the Site wetlands described these areas as a wetland mosaic, with water regimes varying from temporary to saturated, and vegetation types ranging from wet meadow to marsh. It described these hillside wetlands as seep-fed, usually with multiple discharge points located just at or below the pediment surface.

4.2.2 Field Work and Data Analysis

Monitoring was conducted at 15 transects located in the wetlands shown in Figure 4-1. Five 50-m transects were located randomly in each of the wetland areas designated as W1, W2, and W3. Transects were located using the Site's GIS. A baseline was positioned along an edge of the wetland, and randomly generated x and y coordinates were used to mark the 0-m end of the transects on maps. In the field, the 0-m ends of the transects were located on the ground, and the direction of each transect was determined using randomly generated aspects. Adjustments were made as necessary, however, so that each transect remained entirely within the boundaries of the wetland (minimum distance from edge of wetland = 1 m). Tall marsh and short marsh classifications were used as the definition for wetland vegetation for the purposes of this study. A minimum of 3 m was maintained between transects, and no transects overlapped.

The following data were recorded for each transect: species richness, foliar cover, frequency, and Canada thistle and woody plant densities. Each transect was also documented with a photograph looking down the length of the transect. Sampling was conducted only once during the summer (July 13–17, 1998) to prevent trampling and damage to the wetland vegetation along the transects.

- Species richness was determined in a 2-m-wide belt centered along the length of each 50-m transect. All plant species rooted within the

100-m² belt were recorded. In addition, the densities of the woody plant stems and cactus species were counted and recorded for each 100-m² area.

- Foliar cover was estimated using a point-intercept method along each 50-m transect. A 2-m-long rod, with a 6-mm diameter, was dropped vertically at 50-cm intervals along the transect to record a total of 100 intercept points. Foliar vegetation hits (defined as any portion of a plant touching the rod above the ground surface) were recorded in three categories as defined by height and growth form. The topmost hit of each growth form was recorded. The growth forms measured were herbaceous, woody <2 m in height, and woody >2 m in height.
- Frequency information by species was gathered by randomly locating five 1-m² quadrats along the right-hand side (starting from the 0-m end) of each transect and recording all species present in each plot.
- Density counts of Canada thistle (*Cirsium arvense*) were made using a ¼-m² quadrat placed in the bottom left-hand corner of each 1-m² quadrat. The density values made from the smaller quadrats were then multiplied by 4 to provide a 1-m² density value. A single photograph of each transect was taken during the sampling session to visually document the condition of the transect. The photograph was taken from the 0-m end of the transect and looked down the length of the transect toward the 50-m endpoint.

More detailed descriptions of these specific methods are found in the *Environmental Monitoring Department Operating Procedures Manual* (DOE 1995) and the *High Value Vegetation Survey Plan for the Rocky Flats Environmental Technology Site* (K-H 1997a).

Species richness data were summarized by generating a species list for each wetland area. A Sorenson coefficient of similarity index (Brower and Zar 1977) was used to evaluate the species richness similarity among the wetlands. In addition, other species richness variables were calculated from the species lists. Foliar cover data were reported as frequency, absolute cover, and relative cover for each species encountered. Frequency from the cover data was defined as the percent of point-intercept transects in which a species occurred, out of the total possible five transects sampled in each wetland.

Absolute foliar cover was the percentage of the number of hits on a species out of the total number of hits possible at a wetland (500). Relative foliar cover was the number of hits a species had relative to the total number of all vegetative hits recorded per wetland (i.e., the percent of vegetative cover represented by the species). Both absolute and relative foliar cover values are means. Frequency based on quadrats ($n = 25$ per wetland) was defined as the number of quadrats in which a species was recorded, divided by 25 (the total number of quadrats possible) and then multiplied by 100. Density count data were summarized as the mean number of stems per square meter. No statistical analysis of the data was conducted because these data are baseline values, which will be compared with future sampling results.

4.3 Results

Total species richness recorded across all three sites was 95 species. Species richness was highest at site W1 (72 species) and lowest at W2 (54 species; Table 4-1). The highest number of species per 1-m² quadrat was found at site W3 (10.0 species), followed by W1 (8.9 species) and W2 (5.8 species). The percentage of native species was essentially the same at all three sites (68–69 percent; Table 4-1). Sorted by U.S. Fish and Wildlife Service (USFWS) wetland indicator types, obligate wetland species made up the largest number of species at each site (Table 4-1). Of the species recorded at the three sites, none is considered rare or imperiled by the Colorado Natural Heritage Program (CNHP 1997). The Sorenson similarity index showed the greatest similarity (based on species presence/absence) between sites W1 and W3 (0.73). The lowest similarity was between sites W1 and W2 (0.62). Similarity between W2 and W3 was 0.70.

Total absolute foliar cover was essentially equal at sites W1 and W3, with 94.6 and 94 percent, respectively (Table 4-2). Site W2 had only slightly less absolute foliar cover, at 88 percent (Table 4-2). Relative foliar cover was dominated at all sites by Arctic rush (*Juncus balticus*; mean across all three sites = 43.7 percent; Table 4-2). Broad-leaved cattails (*Typha latifolia*) provided the second largest amount of relative foliar cover at sites W1 and W3 (9.3 and 7.7 percent, respectively; Table 4-2). At W2, woolly sedge (*Carex lanuginosa*) provided the second highest amount of relative foliar cover (6.8 percent; Table 4-2). The dominant noxious weed found at all three wetland sites was Canada thistle (*Cirsium arvense*), which averaged 5 percent relative foliar cover across all three sites (Table 4-2). Other species that provided greater than 4 percent relative foliar cover at any of the sites included yellowrocket wintercress (*Barbarea vulgaris*), Nebraska sedge (*Carex nebrascensis*), longstyle rush (*Juncus longistylis*), rough bent (*Agrostis scabra*), Kentucky bluegrass (*Poa pratensis*), and prairie cordgrass (*Spartina pectinata*; Table 4-2). Ninety percent of the foliar cover at W2 came from native species, whereas at sites W1 and W3, only 80 percent of the cover was from native species. Shannon-Weaver diversity indices were calculated for all three sites based on cover data (Table 4-2). Sites W3 (1.07) and W1 (1.03) had the highest diversity, and W2 had the lowest (0.84).

Yellowrocket wintercress had the highest frequency of any recorded species, with frequencies of 100 percent and 92 percent at sites W1 and W3, respectively (Table 4-3). However, it did not occur in any of the quadrats at site W2 (Table 4-3). Arctic rush (mean frequency = 77 percent) and Canada thistle (mean frequency = 75 percent) were the next most common species encountered across all three wetland sites (Table 4-3).

Snowberry (*Symphoricarpos occidentalis*) had the highest woody stem densities of any woody species at all three sites, but was less than 2 stems/m² at all sites (Table 4-4). The noxious weed Canada thistle was found at the highest densities at sites W1 and W3, with plant densities of 13 and 16 plants/m², respectively (Table 4-5). At W2, the Canada thistle density was only 6 plants/m² (Table 4-5).

4.4 Discussion

Overall, the vegetation composition of these three large wetland areas was fairly similar. All three areas were dominated by arctic rush. One of two sedges—either wooly sedge or Nebraska sedge, depending on site location—and cattails provided the remainder of the dominant cover. Differences among the three wetland sites appeared to depend the drainage in which they were located. Sites W1 and W3, in Rock Creek, appeared more similar to one another than to W2, which was located in Woman Creek. Higher total site species richness, number of species/m², total absolute foliar cover, and greater diversity were found at the Rock Creek wetlands. The highest Sorenson similarity index (based on species presence/absence) occurred between the two Rock Creek sites. Data from 1997 species richness inventories of the entire wetland complexes where the transects for this study were located also revealed similar results (K-H 1997c). These data showed that the highest species richness was found at the W1 wetland complex (188 species), and although the lowest species richness occurred at W3 (178 species; W2 had 180 species), the highest similarity occurred between the Rock Creek wetlands (Sorenson coefficient of similarity = 0.80; K-H 1997c).

Compared to the other plant communities monitored on the Site using the same methodology, the total species richness for the wetland community was lower than that observed in the xeric mixed grassland, mesic mixed grassland, and riparian communities² (K-H 1997d). Only the reclaimed grassland had lower species richness. The percentage of native species in the wetland community (overall = 72 percent) was most similar to the riparian community, which would be expected because many of the same species are found in both communities (K-H 1997d). Total foliar cover in the wetlands (92.2 percent) was higher than that found at any of the other plant communities monitored from 1993 through 1995, with the exception of the 1995 mesic mixed grassland (K-H 1997d).

Based on the results of this study, the classification of these large hillside seep wetlands best fits that described by Cooper (1988), in a report on Boulder Valley wetlands, as an arctic rush (*Juncus balticus* = *J. arcticus*) wetland community. Small inclusions of the cattail-duckweed (*Typha latifolia-Lemna minor*), Nebraska sedge, and prairie cordgrass (*Spartina pectinata*) communities are also present within these larger complexes at some locations. With the exception of the prairie cordgrass wetland type, none of these wetland types are uncommon in the greater Boulder area (Cooper 1988). The prairie cordgrass wetland community however, as Cooper (1988) mentions, is more restricted now than in presettlement times. It was previously much more common along the floodplains of rivers and in sloughs and oxbows. As a result, this community is listed by the Colorado Natural Heritage Program (CNHP 1997) as a plant community of concern. It is considered to be a critically imperiled community in the state of Colorado due to its rarity. Although only small patches of the community occur at the Site within the

² The riparian community as used here refers to the EcMP classification system and data sets. It usually included riparian woodland and some small streamside wetland areas.

wetlands, the presence of this wetland type is further evidence of the uniqueness, health, and high quality of the ecological resources, which have been preserved at the Site.

In describing the wetlands of the Boulder valley, Cooper describes the arctic rush community as “occupying seasonally wet meadows.” This classification is described as an herbaceous wetland with mineral soils and fresh water. It is dominated by arctic rush, but often has a variety of associated species. He also mentions that this wetland classification usually has a long grazing history and that arctic rush is considered to be an “increaser,”³ because it is not considered very palatable by cattle. This latter information raises some interesting questions that relate past land use to the current species composition found in Site wetlands. Prior to DOE acquisition in the early 1950s, the entire area of the Site was rangeland that had probably been grazed for at least a century or more (total time frame hypothesized). Considering that arctic rush is an “increaser” with grazing pressure, it would have been increasing in cover for some time in the wetlands on the Site. This could help explain the dominance of arctic rush not only in these wetlands, but many others on the Site. With the relief of grazing pressure from most of these wetlands for nearly 50 years (part of the W3 wetland has not been grazed for only 25 years), it could be assumed that the dominance of arctic rush may be decreasing. The question could also be raised as to whether these wetlands are reverting back to a pre-grazing composition. In other words, as the dominance of arctic rush in the community decreases, are other species migrating back into the community and surviving? Given the prevalence of exotic weed species, especially Canada thistle, it is unknown whether a truly pre-grazing composition could ever be achieved.

A qualitative comparison of the Site 1998 wetland data with information from wetlands on City of Boulder Open Space properties to the north and west of the Site reveals similarities (D’Amico 1998, pers. comm.). Although most of the Open Space wetlands are not large hillside seep wetlands—most are riparian or pond-edge wetlands—the species found at these locations are either present in the 1998 site wetland inventory lists or from the species inventories conducted in wetlands on the Site in 1997. Because of differences in methods, and because the Open Space information was qualitative, no detailed comparisons of species richness or cover were possible. However, the dominant species listed for the Open Space wetlands are also dominant in places on the Site as well. One apparent difference in the Open Space data was the lack of Canada thistle at most localities. Out of 10 wetlands for which data was obtained, Canada thistle was only listed in the species lists for three locations. Whether this is reality or a consequence of the qualitative assessment used to gather the data is not known. If true, it is interesting that these wetlands have not been infested.

On the Site, the noxious weed Canada thistle was found throughout all three wetland sites studied, although it occurred less frequently and had less cover at W2 than at W1 and W3. Canada thistle stem density in Rock Creek (W1 = 13 plants/m²; W3 16 plants/m²)

³ An increaser is defined as a species that increases in dominance because the grazing animals selectively eat other species (i.e., decreasing their abundance), giving the “increaser” species a competitive advantage.

was more than twice that in Woman Creek ($W_2 = 6$ plants/ m^2). Canada thistle is listed on the state noxious weed list as one of the top ten weed species needing control in Colorado (CRS 1996). Additionally, it has been listed by Jefferson County as a priority for control within the county (Lyle 1998). Landowners are responsible for controlling infestations of noxious weeds on their properties and preventing their spread to neighboring landowners.

Control of the Canada thistle in the wetlands on the Site is made difficult because of the open water often present in these areas. Most recommended measures for controlling Canada thistle infestations are designed for dryland infestations, where mowing combined with herbicide treatment can provide effective treatment (Beck 1996). However, within Site wetlands, mowing is not feasible due to the soft, uneven, hummocky ground. Herbicides are not safe to apply at a broad scale because either 1) they are not designed for direct application to water sources, or 2) if they are approved for water application, they are non-selective and would affect all broadleaf forbs in the wetlands. Because none of these options is desirable, alternative solutions must be developed. In addition, repeated application over several years using any means is generally required for effective control of Canada thistle.

Given these conditions, the following control methodology is suggested for controlling Canada thistle in the wetlands on the Site. Two biocontrol insects are available from the Colorado Department of Agriculture (CDA) for control of Canada thistle (Beck 1996). *Ceutorhynchus litura* (crown boring weevil) and *Urophora cardui* (a gall-forming insect) both can stress populations of Canada thistle. *Ceutorhynchus litura* causes plants to be stressed and less vigorous. *Urophora cardui* stresses a plant by causing galls to develop on the plant, which if formed near terminal growing points, prevents flowers from developing and setting seed. Neither is generally capable of totally controlling infestations, but using both in conjunction with other methods has proven effective (Beck 1996). In 1997, the CDA released two biocontrol insects on the Site to assist in the control of diffuse knapweed (*Centaurea diffusa*) and dalmatian toadflax (*Linaria dalmatica*), at no cost to the Site. The CDA should be encouraged to continue to use the Site as a testing ground for releases of the biocontrol insects for Canada thistle. This would provide additional control of this species on Site at little to no cost.

Other potential actions include conducting controlled burns in the wetlands. Although this may not directly reduce or control the Canada thistle, burning would reduce built-up dead plant litter and recycle nutrients, the intent being to invigorate the native plants in the wetlands. This would help the native species to compete with the weeds. Application of approved herbicides is possible in the wetlands using a wick application method, where the herbicide is applied by hand to individual plants. Although more time and labor intensive, this could be done whether controlled burns were conducted first or not. However, because the wetland vegetation is typically dense, removal of the dead plant material from the wetland first would make wick application more effective, as Canada thistle plants and rosettes would be more visible and accessible for herbicide application. Wick applications of herbicides would have to be continued for several years to maintain good control.

4.5 References

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Table 4-1. 1998 wetlands species richness

Family	Scientific Name	Speccode	Native	Site		
				W1	W2	W3
ALISMATACEAE	Sagittaria latifolia Willd.	SALA1	Y			X
APIACEAE	Cicuta maculata L. var. angustifolia Hook.	CIMA1	Y	X		
ASCLEPIADACEAE	Asclepias incarnata L.	ASIN1	Y		X	X
ASCLEPIADACEAE	Asclepias speciosa Torr.	ASSP1	Y	X	X	X
ASTERACEAE	Achillea millefolium L. ssp. lanulosa (Nutt.) Piper	ACMI1	Y	X	X	X
ASTERACEAE	Arnica fulgens Pursh.	ARFU1	Y	X		
ASTERACEAE	Arctium minus Bernh.	ARMI1	Y	X		
ASTERACEAE	Aster falcatus Lindl.	ASFA1	Y	X	X	
ASTERACEAE	Cirsium arvense (L.) Scop.	CIAR1	N	X	X	X
ASTERACEAE	Cirsium vulgare (Savi) Ten.	CIVU1	N	X	X	X
ASTERACEAE	Lactuca serriola L.	LASE1	N	X	X	X
ASTERACEAE	Sonchus asper (L.) Hill	SOAS1	N		X	
ASTERACEAE	Solidago missouriensis Nutt.	SOMI1	Y	X		
ASTERACEAE	Taraxacum officinale Weber	TAOF1	N	X	X	
ASTERACEAE	Tragopogon dubius Scop.	TRDU1	N	X	X	X
BORAGINACEAE	Cynoglossum officinale L.	CYOF1	N		X	X
BORAGINACEAE	Onosmodium molle Michx. var. occidentale (Mack.) Johnst.	ONMO1	Y	X		
BRASSICACEAE	Barbarea vulgaris R. Br.	BAVU1	N	X		X
BRASSICACEAE	Nasturtium officinale R. Br.	NAOF1	N	X		X
BRASSICACEAE	Thlaspi arvense L.	THAR1	N	X	X	X
CANNABACEAE	Humulus lupulus L. var. lupuloides E. Small	HULU1	Y	X		
CAPRIFOLIACEAE	Symporicarpus occidentalis Hook.	SYOC1	Y	X	X	X
CERATOPHYLLACEAE	Ceratophyllum demersum L.	CEDE1	Y			X
CLusiaceae	Hypericum perforatum L.	HYPE1	N	X	X	X
CYPERACEAE	Carex hystericina Muhl. ex Willd.	CAHY1	Y			X
CYPERACEAE	Carex interior Bailey	CAIN1	Y		X	X
CYPERACEAE	Carex lanuginosa Michx.	CALA1	Y	X	X	X
CYPERACEAE	Carex nebrascensis Dew.	CANE1	Y	X	X	X
CYPERACEAE	Carex praegracilis W. Boott.	CAPR1	Y		X	
CYPERACEAE	Carex scoparia Schkuhr. ex Willd.	CASC1	Y	X		
CYPERACEAE	Eleocharis macrostachya Britt.	ELMA1	Y	X	X	X
CYPERACEAE	Scirpus pallidus (Britt.) Fern	SCPA1	Y		X	
EQUISETACEAE	Equisetum laevigatum A. Br.	EQLA1	Y	X	X	
EUPHORBIACEAE	Euphorbia robusta (Engelm.) Small	EURO1	Y			X
FABACEAE	Amorpha fruticosa L.	AMFR1	Y	X		
FABACEAE	Dalea purpurea Vent	DAPU1	Y	X		
FABACEAE	Glycyrrhiza lepidota Pursh.	GLLE1	Y	X		
FABACEAE	Thermopsis rhombifolia var. divaricarpa (Nels.) Isely	THRH1	Y	X		
FABACEAE	Trifolium sp.	TRI1				X
IRIDACEAE	Iris missouriensis Nutt.	IRMI1	Y		X	
JUNCACEAE	Juncus balticus Willd.	JUBA1	Y	X	X	X
JUNCACEAE	Juncus dudleyi Wieg.	JUDU1	Y	X	X	X
JUNCACEAE	Juncus ensifolius Wikst. var. montanus (Engelm.) C. L. Hitchc.	JUEN1	Y		X	X
JUNCACEAE	Juncus longistylis Torr.	JULO1	Y	X	X	X
JUNCACEAE	Juncus nodosus L.	JUNO1	Y		X	X
JUNCACEAE	Juncus torreyi Cov.	JUTO1	Y		X	
LAMIACEAE	Lycopus americanus Muhl. ex Barton	LYAM1	Y	X	X	X
LAMIACEAE	Mentha arvensis L.	MEAR1	Y	X	X	X
LAMIACEAE	Monarda fistulosa L. var. menthifolia (Grah.) Fern.	MOFI1	Y			X
LAMIACEAE	Nepeta cataria L.	NECA1	N	X		X
LAMIACEAE	Prunella vulgaris L.	PRVU1	Y	X	X	X
LEMNACEAE	Lemna minor L.	LEMI1	Y	X	X	X
LILIACEAE	Allium textile A. Nels. & Macbr.	ALTE1	Y	X		
LILIACEAE	Smilacina stellata (L.) Desf.	SMST1	Y	X		
LYTHRACEAE	Lythrum alatum Pursh.	LYAL1	Y		X	

Table 4-1. (cont.)

Family	Scientific Name	Speccode	Native	Site		
				W1	W2	W3
ONAGRACEAE	<i>Epilobium ciliatum</i> Raf. ssp. <i>glandulosum</i> (Lehm.) Hock & Raven	EPCI1	Y	X	X	X
ONAGRACEAE	<i>Epilobium paniculatum</i> Nutt.	EPPA1	Y	X	X	X
ONAGRACEAE	<i>Oenothera villosa</i> Thunb. ssp. <i>strigosa</i> (Rydb.) Dietrich & Raven	OEV1	Y	X	X	X
OXALIDACEAE	<i>Oxalis dillenii</i> Jacq.	OXDI1	N	X		X
POACEAE	<i>Agropyron repens</i> (L.) Beauv.	AGRE1	N	X	X	X
POACEAE	<i>Agrostis scabra</i> Willd.	AGSC1	Y	X	X	X
POACEAE	<i>Agrostis stolonifera</i> L.	AGST1	N	X	X	X
POACEAE	<i>Bromus inermis</i> Leyss. ssp. <i>inermis</i>	BRIN1	N	X		
POACEAE	<i>Bromus japonicus</i> Thunb. ex Murr.	BRJA1	N	X	X	X
POACEAE	<i>Calamagrostis stricta</i> (Timm.) Koel	CAST2	Y		X	
POACEAE	<i>Glyceria striata</i> (Lam.) Hitchc.	GLST1	Y	X	X	X
POACEAE	<i>Hordeum jubatum</i> L.	HOJU1	Y	X		X
POACEAE	<i>Poa compressa</i> L.	POCO1	N		X	
POACEAE	<i>Poa palustris</i> L.	POPA1	N	X	X	X
POACEAE	<i>Poa pratensis</i> L.	POPR1	N	X	X	X
POACEAE	<i>Sphenopholis obtusata</i> (Michx.) Scribn.	SPOB1	Y		X	X
POACEAE	<i>Spartina pectinata</i> Link	SPPE1	Y	X	X	X
POLYGONACEAE	<i>Polygonum convolvulus</i> L.	POCO2	N	X		X
POLYGONACEAE	<i>Polygonum ramosissimum</i> Michx.	PORA1	Y	X		X
POLYGONACEAE	<i>Rumex crispus</i> L.	RUCR1	N	X	X	X
POLYGONACEAE	<i>Rumex obtusifolius</i> L.	RUOB1	N	X		
PRIMULACEAE	<i>Lysimachia ciliata</i> L.	LYCI1	Y	X		
RANUNCULACEAE	<i>Ranunculus macounii</i> Britt.	RAMA1	Y	X		X
ROSACEAE	<i>Geum aleppicum</i> Jacq.	GEAL1	Y	X	X	X
ROSACEAE	<i>Geum macrophyllum</i> Willd.	GEMA1	Y	X	X	X
ROSACEAE	<i>Potentilla gracilis</i> Dougl. ex Hook. var. <i>glabrata</i> (Lehm.) C. L. Hitchc.	POGR1	Y	X	X	
ROSACEAE	<i>Potentilla norvegica</i> L.	PONO1	Y	X		X
ROSACEAE	<i>Rosa arkansana</i> Porter	ROAR1	Y	X		X
RUBIACEAE	<i>Galium aparine</i> L.	GAAP1	Y	X		X
RUBIACEAE	<i>Galium septentrionale</i> Roemer & Schultes	GASE1	Y	X		
SCROPHULARIACEAE	<i>Mimulus glabratus</i> H. B. K. var. <i>fremontii</i> (Benth.) A. L. Grant	MIGL1	Y			X
SCROPHULARIACEAE	<i>Scrophularia lanceolata</i> Pursh.	SCLA2	Y	X		
SCROPHULARIACEAE	<i>Veronica anagallis-aquatica</i> L.	VEAN1	N	X		X
SCROPHULARIACEAE	<i>Verbascum blattaria</i> L.	VEBL1	N		X	
SCROPHULARIACEAE	<i>Verbascum thapsus</i> L.	VETH1	N	X		X
TYPHACEAE	<i>Typha latifolia</i> L.	TYLA1	Y	X	X	X
UNKNOWN	Unidentifiable species	UNKN		X		
URTICACEAE	<i>Urtica dioica</i> L. ssp. <i>gracilis</i> (Ait.) Seland.	URDI1	Y	X		X
VERBENACEAE	<i>Verbena hastata</i> L.	VEHA1	Y	X	X	X
VIOLACEAE	<i>Viola sororia</i> Willd.	VISO1	Y			X
Total # species:				72	54	63
Percent native species:				69	69	68

Wetland Indicator Species

Facultative species	FAC	14	6	9
Facultative upland species	FACU	15	9	1
Facultative upland species - less frequently found in wetlands	FACU-	0	0	1
Facultative wetland species	FACW	12	12	10
Non-indicator species	NI	9	8	8
Obligate wetland species	OBL	14	15	19
Upland species	UPL	1	2	1
Total # species:		65	52	49

Table 4-2. 1998 wetland foliar cover data summary

Family	Scientific Name	Speccode	W1			W2			W3		
			Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover
ASCLEPIADACEAE	<i>Asclepias incarnata</i> L.	ASIN1							80	3.60	3.83
ASCLEPIADACEAE	<i>Asclepias speciosa</i> Torr.	ASSP1	40	1.40	1.48	20	0.20	0.23			
ASTERACEAE	<i>Achillea millefolium</i> L. ssp. <i>lanulosa</i> (Nutt.) Piper	ACMI1	20	0.20	0.21						
ASTERACEAE	<i>Cirsium arvense</i> (L.) Scop.	CIAR1	80	5.40	5.71	40	3.40	3.86	100	5.20	5.53
ASTERACEAE	<i>Lactuca serriola</i> L.	LASE1							20	0.20	0.21
BRASSICACEAE	<i>Barbarea vulgaris</i> R. Br.	BAVU1	80	7.40	7.82				100	4.60	4.89
BRASSICACEAE	<i>Nasturtium officinale</i> R. Br.	NAOF1	20	0.80	0.85				20	3.20	3.40
CLUSIACEAE	<i>Hypericum perforatum</i> L.	HYPE1	20	1.40	1.48	20	0.20	0.23			
CYPERACEAE	<i>Carex hystericina</i> Muhl. ex Willd.	CAHY1							20	0.20	0.21
CYPERACEAE	<i>Carex interior</i> Bailey	CAIN1							40	0.40	0.43
CYPERACEAE	<i>Carex lanuginosa</i> Michx.	CALA1	60	7.60	8.03	80	6.00	6.82	20	0.40	0.43
CYPERACEAE	<i>Carex nebrascensis</i> Dew.	CANE1	80	3.00	3.17	80	5.00	5.68	100	6.00	6.38
CYPERACEAE	<i>Carex praegracilis</i> W. Boott.	CAPR1				60	1.60	1.82			
CYPERACEAE	<i>Eleocharis macrostachya</i> Britt.	ELMA1	20	0.80	0.85	20	0.80	0.91	80	2.80	2.98
FABACEAE	<i>Thermopsis rhombifolia</i> var. <i>divaricarpa</i> (Nels.) Isely	THRH1	40	1.20	1.27						
JUNCACEAE	<i>Juncus balticus</i> Willd.	JUBA1	100	37.80	39.96	100	45.20	51.36	100	37.40	39.79
JUNCACEAE	<i>Juncus ensifolius</i> Wikst. var. <i>montanus</i> (Engelm.) C. L. Hitchc.	JUEN1				20	0.20	0.23	20	0.20	0.21
JUNCACEAE	<i>Juncus longistylis</i> Torr.	JUL01	40	2.20	2.33	40	3.80	4.32	60	1.60	1.70
JUNCACEAE	<i>Juncus nodosus</i> L.	JUNO1				20	0.40	0.45			
LAMIACEAE	<i>Mentha arvensis</i> L.	MEAR1	80	3.40	3.59				40	1.80	1.91
LAMIACEAE	<i>Monarda fistulosa</i> L. var. <i>menthifolia</i> (Grah.) Fern.	MOFI1							20	0.20	0.21
LAMIACEAE	<i>Nepeta cataria</i> L.	NECA1							20	0.20	0.21
LAMIACEAE	<i>Prunella vulgaris</i> L.	PRVU1				20	0.20	0.23			
LEMNACEAE	<i>Lemna minor</i> L.	LEMI1				20	0.20	0.23	40	0.40	0.43
LILIACEAE	<i>Smilacina stellata</i> (L.) Desf.	SMST1	20	0.20	0.21						
LYTHRACEAE	<i>Lythrum alatum</i> Pursh.	LYAL1				20	0.40	0.45			
ONAGRACEAE	<i>Epilobium ciliatum</i> Raf. ssp. <i>glandulosum</i> (Lehm.) Hock & Raven	EPC11	60	1.20	1.27	20	0.20	0.23	40	0.40	0.43
ONAGRACEAE	<i>Epilobium paniculatum</i> Nutt.	EPPA1	40	0.60	0.63	20	0.40	0.45	80	1.40	1.49
ONAGRACEAE	<i>Oenothera villosa</i> Thunb. ssp. <i>strigosa</i> (Rydb.) Dietrich & Raven	OEV11	20	0.20	0.21				40	0.80	0.85
POACEAE	<i>Agropyron repens</i> (L.) Beauv.	AGRE1	40	0.40	0.42	20	0.40	0.45			
POACEAE	<i>Agrostis scabra</i> Willd.	AGSC1	20	0.40	0.42				80	4.60	4.89
POACEAE	<i>Agrostis stolonifera</i> L.	AGST1	40	0.60	0.63				60	1.60	1.70
POACEAE	<i>Calamagrostis stricta</i> (Timm.) Koel	CAST2				20	1.60	1.82			
POACEAE	<i>Glyceria striata</i> (Lam.) Hitchc.	GLST1							20	1.20	1.28
POACEAE	<i>Hordeum jubatum</i> L.	HOJU1	20	0.20	0.21				20	0.20	0.21
POACEAE	<i>Poa pratensis</i> L.	POPR1	60	2.40	2.54	40	4.40	5.00	80	3.20	3.40
POACEAE	<i>Spartina pectinata</i> Link	SPPE1	20	1.80	1.90	40	5.00	5.68	40	1.40	1.49
POACEAE	<i>Sphenopholis obtusata</i> (Michx.) Scribn.	SPOB1							20	0.20	0.21

Table 4-2. (cont.)

Family	Scientific Name	Speccode	W1			W2			W3		
			Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover
POLYGONACEAE	<i>Polygonum convolvulus</i> L.	POCO2	20	0.40	0.42				20	0.40	0.43
POLYGONACEAE	<i>Rumex crispus</i> L.	RUCR1				20	0.20	0.23			
ROSACEAE	<i>Geum aleppicum</i> Jacq.	GEAL1	20	0.20	0.21	60	1.40	1.59	40	0.40	0.43
ROSACEAE	<i>Geum macrophyllum</i> Willd.	GEMA1	80	2.60	2.75	80	1.00	1.14	60	1.60	1.70
RUBIACEAE	<i>Galium aparine</i> L.	GAAP1	20	0.40	0.42				20	0.20	0.21
RUBIACEAE	<i>Galium septentrionale</i> Roemer & Schultes	GASE1	20	0.40	0.42						
SCROPHULARIACEAE	<i>Scrophularia lanceolata</i> Pursh.	SCLA2	20	0.20	0.21						
SCROPHULARIACEAE	<i>Verbascum thapsus</i> L.	VETH1	20	0.20	0.21						
SCROPHULARIACEAE	<i>Veronica anagallis-aquatica</i> L.	VEAN1							20	0.20	0.21
TYPHACEAE	<i>Typha latifolia</i> L.	TYLA1	60	8.80	9.30	40	5.80	6.59	40	7.20	7.66
URTICACEAE	<i>Urtica dioica</i> L. ssp. <i>gracilis</i> (Ait.) Seland.	URDI1							20	0.20	0.21
VERBENACEAE	<i>Verbena hastata</i> L.	VEHA1	20	0.60	0.63				20	0.20	0.21
VIOLACEAE	<i>Viola sororia</i> Willd.	VISO1							20	0.20	0.21
UNKNOWN	Unknown species	UNKN	20	0.20	0.21						
Total cover				94.60	100.00		88.00	100.00		94.00	100.00
Shannon-Weiner diversity index				1.03			0.84			1.07	

Note: Absolute cover = Absolute foliar cover is the percentage of the number of hits on a species out of the total number of hits possible at a wetland (500).

Relative cover = Relative foliar cover is the number of hits on a species relative to the total number of all vegetative hits recorded per wetland (i.e., the percent of vegetative cover the species represented).

All cover values presented are means (n = 5).

Table 4-3 1998 wetland species frequency data summary

Family	Scientific Name	Speccode	Native	Site		
				W1 Frequency	W2 Frequency	W3 Frequency
APIACEAE	Cicuta maculata L. var. angustifolia Hook.	CIMA1	Y	4		
ASCLEPIADACEAE	Asclepias incarnata L.	ASIN1	Y			32
ASCLEPIADACEAE	Asclepias speciosa Torr.	ASSP1	Y	8	4	
ASTERACEAE	Achillea millefolium L. ssp. lanulosa (Nutt.) Piper	ACMI1	Y	20	4	4
ASTERACEAE	Arnica fulgens Pursh.	ARFU1	Y	4		
ASTERACEAE	Aster falcatus Lindl.	ASFA1	Y	12		
ASTERACEAE	Cirsium arvense (L.) Scop.	CIAR1	N	80	56	88
ASTERACEAE	Cirsium vulgare (Savi) Ten.	CIVU1	N	12		8
ASTERACEAE	Lactuca serriola L.	LASE1	N	4		16
ASTERACEAE	Taraxacum officinale Weber	TAOF1	N	12		
BORAGINACEAE	Cynoglossum officinale L.	CYOF1	N		4	
BORAGINACEAE	Onosmodium molle Michx. var. occidentale (Mack.) Johnst.	ONMO1	Y	4		
BRASSICACEAE	Barbarea vulgaris R. Br.	BAVU1	N	100		92
BRASSICACEAE	Nasturtium officinale R. Br.	NAOF1	N	4		16
BRASSICACEAE	Thlaspi arvense L.	THAR1	N	12		
CAPRIFOLIACEAE	Syphoricarpos occidentalis Hook.	SYOC1	Y	12	32	20
CLusiaceae	Hypericum perforatum L.	HYPE1	N	16	4	12
CYPERACEAE	Carex interior Bailey	CAIN1	Y			4
CYPERACEAE	Carex lanuginosa Michx.	CALA1	Y	20	20	
CYPERACEAE	Carex nebrascensis Dew.	CANE1	Y	20	24	20
CYPERACEAE	Carex praegracilis W. Boott.	CAPR1	Y		4	
CYPERACEAE	Carex scoparia Schkuhr. ex Willd.	CASC1	Y	4		
CYPERACEAE	Eleocharis macrostachya Britt.	ELMA1	Y	4	8	16
FABACEAE	Trifolium sp.	TRI1				4
JUNCACEAE	Juncus balticus Willd.	JUBA1	Y	72	84	76
JUNCACEAE	Juncus dudleyi Wieg.	JUDU1	Y	4		
JUNCACEAE	Juncus ensifolius Wikst. var. montanus (Engelm.) C. L. Hitchc.	JUEN1	Y			4
JUNCACEAE	Juncus longistylis Torr.	JULO1	Y	8	32	36
JUNCACEAE	Juncus nodosus L.	JUNO1	Y			8
LAMIACEAE	Lycopus americanus Muhl. ex Barton	LYAM1	Y	4		28
LAMIACEAE	Mentha arvensis L.	MEAR1	Y	68	20	44
LAMIACEAE	Monarda fistulosa L. var. menthifolia (Grah.) Fern.	MOFI1	Y			4
LAMIACEAE	Nepeta cataria L.	NECA1	N	4		4
LAMIACEAE	Prunella vulgaris L.	PRVU1	Y	4	4	
LEMNACEAE	Lemna minor L.	LEMI1	Y	4	12	12
LYTHRACEAE	Lythrum alatum Pursh.	LYAL1	Y		20	
ONAGRACEAE	Epilobium ciliatum Raf. ssp. glandulosum (Lehm.) Hock & Raven	EPCI1	Y	44	36	36
ONAGRACEAE	Epilobium paniculatum Nutt.	EPPA1	Y	40	20	56
ONAGRACEAE	Oenothera villosa Thunb. ssp. strigosa (Rydb.) Dietrich & Raven	OEVI1	Y	24	20	16

Table 4-3. (cont.)

Family	Scientific Name	Speccode	Native	Site		
				W1 Frequency	W2 Frequency	W3 Frequency
OXALIDACEAE	<i>Oxalis dillenii</i> Jacq.	OXDI1	N	8		
POACEAE	<i>Agropyron repens</i> (L.) Beauv.	AGRE1	N	4	12	
POACEAE	<i>Agrostis scabra</i> Willd.	AGSC1	Y	16		40
POACEAE	<i>Agrostis stolonifera</i> L.	AGST1	N	8		24
POACEAE	<i>Calamagrostis stricta</i> (Timm.) Koel	CAST2	Y		8	
POACEAE	<i>Glyceria striata</i> (Lam.) Hitchc.	GLST1	Y			8
POACEAE	<i>Hordeum jubatum</i> L.	HOJU1	Y	4		4
POACEAE	<i>Poa palustris</i> L.	POPA1	N		8	
POACEAE	<i>Poa pratensis</i> L.	POPR1	N	32	24	44
POACEAE	<i>Spartina pectinata</i> Link	SPPE1	Y	8	12	4
POACEAE	<i>Sphenopholis obtusata</i> (Michx.) Scribn.	SPOB1	Y			4
POLYGONACEAE	<i>Polygonum convolvulus</i> L.	POCO2	N	4		4
POLYGONACEAE	<i>Polygonum ramosissimum</i> Michx.	PORA1	Y	4		
POLYGONACEAE	<i>Rumex crispus</i> L.	RUCR1	N	4	4	
RANUNCULACEAE	<i>Ranunculus macounii</i> Britt.	RAMA1	Y	4		
ROSACEAE	<i>Geum aleppicum</i> Jacq.	GEAL1	Y	12	36	8
ROSACEAE	<i>Geum macrophyllum</i> Willd.	GEMA1	Y	60	40	56
ROSACEAE	<i>Potentilla gracilis</i> Dougl. ex Hook. var. <i>glabrata</i> (Lehm.) C. L. Hitchc.	POGR1	Y	4		
ROSACEAE	<i>Potentilla norvegica</i> L.	PONO1	Y	4		
ROSACEAE	<i>Rosa arkansana</i> Porter	ROAR1	Y			4
RUBIACEAE	<i>Galium aparine</i> L.	GAAP1	Y	32		56
RUBIACEAE	<i>Galium septentrimoniale</i> Roemer & Schultes	GASE1	Y	8		
SCROPHULARIACEAE	<i>Scrophularia lanceolata</i> Pursh.	SCLA2	Y	16		
SCROPHULARIACEAE	<i>Veronica anagallis-aquatica</i> L.	VEAN1	N			16
TYPHACEAE	<i>Typha latifolia</i> L.	TYLA1	Y	24	16	32
URTICACEAE	<i>Urtica dioica</i> L. ssp. <i>gracilis</i> (Ait.) Seland.	URDI1	Y			8
VERBENACEAE	<i>Verbena hastata</i> L.	VEHA1	Y	4	8	8
VIOLACEAE	<i>Viola sororia</i> Willd.	VISO1	Y			24

Table 4-4. 1998 wetland shrub densities

Scientific Name	Site		
	W1	W2	W3
<i>Amorpha fruticosa L.</i>	0.002	0.000	0.000
<i>Rosa arkansana Porter</i>	0.046	0.000	0.064
<i>Symporicarpos occidentalis Hook.</i>	0.844	1.850	0.822

Note: Values are # stems/sq. meter.

**Table 4-5. 1998 wetland Canada thistle
(*Cirsium arvense*) densities**

Site	Density (# plants/m ²)
W1	12.96
W2	5.76
W3	16.00

Report 5

1998 Xeric Mixed Grassland Monitoring Summary

5. 1998 Xeric Mixed Grassland Monitoring Summary

5.1 Purpose

Monitoring is an integral part of determining whether the management objectives and goals for the high-value plant communities at Rocky Flats Environmental Technology Site (Site) are being achieved (K-H 1997a,b). Consistent with this goal, long-term quantitative monitoring is necessary to determine whether changes are taking place in the plant communities that would go undetected through the use of broader scale qualitative monitoring techniques.

During 1998, three permanent monitoring sites (TR01, TR06, and TR12) in the xeric mixed grassland community that had been set out and monitored in 1993, 1994, and 1995 (Figure 5-1; DOE 1995a, K-H 1997c) were monitored again to reassess and document any change. Originally, all three sites were classified as xeric mixed grassland. After the 1996 vegetation mapping effort, sites TR01 and TR12 were reclassified as part of the xeric tallgrass prairie community, and TR06 was reclassified as part of the xeric needle and threadgrass community, to better reflect the actual community composition of the xeric mixed grassland (K-H 1997c). These sites were last monitored in the summer of 1995. The purpose of this year's monitoring was to re-evaluate the health of the plant communities at these locations and document any change.

5.2 Background Information

The plant communities monitored from 1993 through 1995 were organized along a soil moisture (hydrologic) gradient that ranged from xeric (dry) to mesic (moderate moisture) to hydric (wet). This classification followed the plant community classification that had been outlined in the baseline study (DOE 1992), which identified xeric (xeric mixed grassland), mesic (mesic mixed grassland), and hydric (riparian community) communities at the Site. Since the last time these sites were monitored in 1995, some weed control efforts had been conducted at some locations. In June 1997, TR12 was sprayed with Tordon 22K to control the noxious weed diffuse knapweed (*Centaurea diffusa*), and in August 1997, one of the transects at TR06 had a biological control agent released to help control another weed, dalmatian toadflax (*Linaria dalmatica*), which is abundant at that location.

5.3 Methods

During 1998, the xeric sites, TR01, TR06, and TR12 (Figure 5-1), were monitored for species richness, cover, and frequency. The sampling methods and procedures used at these sites during 1998 were the same as those used in 1993–1995, and are described in the *Ecological Monitoring Program, Final Program Plan* (DOE 1993) and the

Environmental Management Operating Procedures Manual, Volume V, Ecology, 5-51200-OPS-EE (DOE 1995b). An additional measure for species frequency was added to the sampling in 1998 to provide additional quantitative information (described below).

A total of fifteen 50-m transects (five at each site) were monitored in 1998. Transects were sampled in the spring and late summer. Species richness and frequency were monitored during both sampling sessions, and cover was sampled only during the late-summer session. Species richness was determined in a 2-m-wide belt centered along the length of each 50-m transect. Every plant species rooted within the 100-m² area was recorded. In addition, the densities of the woody plant stems and cactus species were counted and recorded for the 100-m² area. Basal cover and foliar cover were estimated using a point-intercept method along each 50-m transect. A 2-m-long rod, with a 6-mm diameter, was dropped vertically at 50-cm intervals along the transect to record a total of 100 intercept points.

Two types of hits were recorded. Basal cover hits were recorded based on what material was hit by the rod at the ground surface. Hits could be vegetation (live plants), litter (fallen dead material), rock (pebbles and cobbles that were greater than the rod diameter), bare ground, or water, in that order of priority based on the protection from erosion provided by each type of cover. Basal vegetation hits were recorded only if the rod was touching the stem or crown of the plant where the plant entered the ground. Foliar vegetation hits (defined as a portion of a plant touching the rod) were recorded in three categories as defined by height and growth form. The topmost hit of each growth form was recorded. The growth forms measured were herbaceous, woody <2 m in height, and woody >2 m in height. Frequency information by species was gathered by randomly locating 25 1-m² quadrats (five per transect) at each site. Additionally, a single photograph of each transect was taken during the late summer sampling session to visually document the condition of the transect. Photographs were taken from the 0-m end of the transect near the permanent marker, looking toward the 50-m endpoint.

Species richness data were summarized by generating a species list for each site. To make the 1998 data compatible with the way past data sets had been analyzed, belt-transect data and point-intercept data were combined to provide overall species richness for analysis. Other species richness variables were calculated from the species lists and used for comparison. Basal cover data are reported as total percent cover of vegetation, litter, rock, and bare ground. Foliar cover data are reported as frequency, absolute cover, and relative cover for each species encountered. Frequency from the cover data was defined as the percent of point-intercept transects on which a species occurred, out of the total possible five sampled at each site. Absolute foliar cover was the percentage of the number of hits on a species out of the total number of hits possible at a site (500). This value is the actual cover of a species. Relative foliar cover was the number of hits a species had relative to the total number of vegetative hits recorded per site (i.e., the percent of total vegetative cover (100 percent) the species represented). Both absolute and relative foliar cover values presented are means. Frequency based on quadrats (n=25) was defined as the number of quadrats in which a species was recorded, divided by 25 (the total number of quadrats possible), and multiplied by 100. Descriptive

comparisons were made between the 1993–1995 and 1998 data sets to examine potential changes over time. No statistical analyses were conducted because of the variability in the data and the short-term nature of the data sets.

5.4 Results

A total of 122 species were recorded at all three sites monitored in 1998. The number of species found at each site varied from 81 to 84, with the site TR01 having the lowest and TR12 the highest (Table 5-1). The percentage of native species found across all sites combined was 84 percent, with individual sites ranging from 81 to 86 percent (Table 5-1). A Sorenson similarity index using species presence/absence data revealed the highest similarity between TR01 and TR12 (0.79). Comparisons between TR06 and TR01, and TR06 and TR12, were lower, at 0.63 and 0.67, respectively.

Cactus density was highest at site TR12 (1.16 plants/m²), followed by TR01 (0.68 plants/m²) and TR06 (0.11 plants/m²). Spanish bayonet (*Yucca glauca*), was the only woody plant of any abundance to occur at any of the sites, and it was only found at TR06 (0.27 plants/m²).

Basal vegetation cover averaged 7–8 percent at all three sites (Table 5-2). Other ground cover classifications, in descending order of importance at all sites, came from litter, rock, and bare ground (Table 5-2). Total foliar cover was similar at all sites, ranging from 81 to 85 percent (Table 5-2). The percentage of cover coming from native vegetation, however, was highest at sites TR01 (84 percent) and TR12 (87 percent; Table 5-2). Native cover was much lower at TR06 (66 percent; Table 5-2). Examined by cool-season vs. warm-season graminoid species, TR01 was the only site dominated by warm-season grasses (41 percent; Table 5-2). Site TR06 was dominated by cool-season graminoid species (80 percent), with warm-season graminoid species constituting only 5 percent of the total vegetation cover (Table 5-2). Site TR12 was intermediate, with cool- and warm-season grasses constituting 63 and 28 percent of the total vegetation cover, respectively (Table 5-2). Site TR01 was dominated by mountain muhly (*Muhlenbergia montana*), Porter's aster (*Aster porteri*), and big bluestem (*Andropogon gerardii*; Table 5-3). At site TR06, the dominant species were needle and thread grass (*Stipa comata*), dalmatian toadflax (*Linaria dalmatica*), and Japanese brome (*Bromus japonicus*; Table 5-3). Site TR12 had foliar cover dominated by needle and thread grass, big bluestem, and Canada bluegrass (*Poa compressa*; Table 5-3). Species frequency results are presented for the first time for both spring and summer sampling sessions in Table 5-4. Different sites had differing frequencies for the various species, and this information will be most useful for comparisons to future monitoring to determine whether there have been any changes.

5.5 Discussion

The permanent transects at sites TR01, TR06, and TR12 were monitored during 1998, and the data were compared to those from 1993–1995. No major changes were observed

for most of the endpoints considered (Table 5-2). In general, most of the measured variables in 1998 were intermediate in comparison to past measured variables, indicating the natural variability inherent in the grassland ecosystem, although some of the variability is likely due to sampling bias as well. For example, the apparent losses of basal vegetation cover in 1998 (which were offset by large increases in litter cover) are most likely due to differences in field technicians' interpretations of basal vegetation and litter hits (Table 5-2).

No important changes were noted with respect to species richness or total foliar cover at any of the sites since they were last monitored. Previously, a loss of relative native cover at these sites had been shown in the data (from 1993–1995; K-H 1997c) and was an issue of concern. The 1998 data showed that this apparent trend may be starting to reverse at sites TR06 and TR12. At TR01, however, native cover continues to decline (Table 5-3). Most of the non-native cover at TR01 in 1998 was from the cool-season graminiod species, Canada bluegrass and Kentucky bluegrass (*Poa pratensis*), both of which showed increased relative foliar cover amounts since 1995 (Table 5-3). Although the decline in overall non-native cover observed at TR01 since 1995 was minimal (2.6 percent), the continuing loss of native foliar cover at this site warrants further observation in future years when these sites are monitored again.

Relative foliar cover values were examined for all species at each site for 1993, 1994, 1995, and 1998, to determine whether any large or consistent changes in relative cover had occurred. A few individual species showed some apparent change worthy of noting. One of the more important species on the xeric tallgrass prairie that has been losing cover over the past few years is little bluestem (*Andropogon scoparius*), one of the relict tallgrass prairie species (Table 5-5). Little bluestem showed steady declines in relative cover at both TR01 and TR12 over the past several years, declining from almost 12 percent to 2 percent at TR01 and 5 percent to 0 percent at TR12 (Table 5-5). Qualitative observations on the loss of little bluestem cover on the Site have been mentioned in the past (K-H 1997c), and now the quantitative measurements confirm this observation. Originally, a large die-off of the species at these and other locations on the Site was noticed in 1995, the year following the late-summer drought of 1994. Little bluestem was particularly hard hit, and observations showed that many of the bunches died. Only time will tell whether the species will recover at these sites. Qualitative observations at some other locations on the Site have shown, however, that the species was apparently not as severely affected and appears to be doing fine. So this may simply be an example of the dynamic nature of these native plant communities in response to environmental changes at different scales.

During 1998, at TR01, mountain muhly cover increased dramatically from past measurements (Table 5-5). Porters' aster at TR01 lost over 10 percent of its relative foliar cover since 1995, when it had a bumper-crop year and was abundant across the prairie (Table 5-5). Whether the large fluctuations in these two species indicate a species composition shift or just natural variability is not certain, however, because the time frames represented by the data sets are too short to separate natural variability, or "noise," from real trends. Data sets of more than 10 years might begin to show some of the

annual variation that could be expected. The apparent increase of Japanese brome (cheatgrass) at TR06 is also of concern, because this site already has the most non-native composition of the three sites (Table 5-5). This warrants continued observation as well.

Site TR12 was treated with the herbicide Tordon 22K in June 1997 to help control diffuse knapweed, which had begun to seriously infest the site. Using 1993–1995 data as baseline information and sites TR01 and TR06 as controls, a few changes have occurred, likely attributable to the herbicide application. The percent of native foliar cover at TR12 rose by 10 percent from 1995 to 1998, compared to continued loss of native cover at TR01 and only a 3 percent increase at TR06 (Table 5-2). Forb cover also decreased by over 20 percent at TR12, compared to 10 percent declines at TR01 and TR06 for the same time period (Table 5-2). These responses are similar to those observed at the diffuse knapweed herbicide monitoring plots for 1998 (see Section 3 in this Annual Report).

Examination of data from the transect at TR06, where biological control agents were released in 1997, showed no observable effect on the cover of dalmatian toadflax. Qualitative visual observations in the area where the agents were released also did not show any noticeable impact on the species. It may take several years for the biological control agents to increase to levels that can cause noticeable impacts on dalmatian toadflax.

In general, the 1998 monitoring data showed that no major changes have occurred at these sites since they were last monitored. The few changes that were noted are subject to the short-term nature of the available data sets, making it difficult to distinguish natural variability “noise” from real trends. However, these data, combined with qualitative assessments of the resources, continue to suggest that proactive management of the grassland communities on the Site is necessary to maintain the quality and health of these communities, and to preserve these resources for future generations. For example, if site TR12 had not been treated with herbicide to control the diffuse knapweed infestation—which qualitative assessments had indicated was a problem during the years since the last quantitative monitoring at the site—the results might have been quite different. Currently, the most significant threat to the grassland communities on the Site is from noxious weeds. The buildup of dead plant litter in the communities, resulting from a lack of fire and grazing, is also a significant problem. As good stewards of the ecological resources at the Site, the implementation of prescribed burns and continued use of various weed control methods (potentially including aerial herbicide application) will provide essential tools for proper management of these resources.

5.6 References

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Table 5-1. 1998 xeric mixed grassland species richness

Family	Scientific Name	Speccode	Native	TR01	TR06	TR12
AGAVACEAE	<i>Yucca glauca</i> Nutt.	YUGL1	Y		X	
ANACARDIACEAE	<i>Rhus aromatica</i> Ait. var. <i>trilobata</i> (Nutt.) A. Gray	RHAR1	Y	X		
APIACEAE	<i>Lomatium orientale</i> Coulter. & Rose	LOOR1	Y	X	X	X
ASCLEPIADACEAE	<i>Asclepias viridiflora</i> Raf.	ASVI1	Y	X		X
ASTERACEAE	<i>Achillea millefolium</i> L. ssp. <i>lanulosa</i> (Nutt.) Piper	ACMI1	Y	X		X
ASTERACEAE	<i>Ambrosia psilostachya</i> DC.	AMPS1	Y	X	X	X
ASTERACEAE	<i>Antennaria parvifolia</i> Nutt.	ANPA1	Y	X		X
ASTERACEAE	<i>Artemisia campestris</i> L. ssp. <i>caudata</i> (Michx.) Hall & Clem.	ARCA1	Y		X	
ASTERACEAE	<i>Artemisia dracunculus</i> L.	ARDR1	Y		X	
ASTERACEAE	<i>Artemisia frigida</i> Willd.	ARFR1	Y	X	X	X
ASTERACEAE	<i>Artemisia ludoviciana</i> Nutt. var. <i>ludoviciana</i>	ARLU1	Y	X	X	X
ASTERACEAE	<i>Aster falcatus</i> Lindl.	ASFA1	Y	X		
ASTERACEAE	<i>Aster porteri</i> Gray	ASPO1	Y	X		X
ASTERACEAE	<i>Carduus nutans</i> L. ssp. <i>macrolepis</i> (Peterm.) Kazmi	CANU1	N		X	
ASTERACEAE	<i>Centaurea diffusa</i> Lam.	CEDI1	N	X		X
ASTERACEAE	<i>Chrysopsis fulcrata</i> Greene	CHFU1	Y	X		X
ASTERACEAE	<i>Chrysopsis villosa</i> Pursh.	CHV11	Y	X	X	X
ASTERACEAE	<i>Cirsium arvense</i> (L.) Scop.	CIAR1	N		X	
ASTERACEAE	<i>Cirsium undulatum</i> (Nutt.) Spreng.	CIUN1	Y	X	X	
ASTERACEAE	<i>Erigeron canus</i> A. Gray	ERCA1	Y		X	
ASTERACEAE	<i>Erigeron divergens</i> T. & G.	ERDI1	Y	X	X	X
ASTERACEAE	<i>Erigeron flagellaris</i> A. Gray	ERFL1	Y			X
ASTERACEAE	<i>Gaillardia aristata</i> Pursh.	GAAR1	Y	X		X
ASTERACEAE	<i>Gutierrezia sarothrae</i> (Pursh.) Britt. & Rusby	GUSA1	Y			X
ASTERACEAE	<i>Helianthus pumilus</i> Nutt.	HEPU1	Y	X	X	
ASTERACEAE	<i>Lactuca serriola</i> L.	LASE1	N	X	X	X
ASTERACEAE	<i>Liatris punctata</i> Hook.	LIPU1	Y	X	X	X
ASTERACEAE	<i>Microseris cuspidata</i> (Pursh.) Sch. Bip.	MICU1	Y	X	X	X
ASTERACEAE	<i>Ratibida columnifera</i> (Nutt.) Woot. & Standl.	RACO1	Y	X		X
ASTERACEAE	<i>Scorzonera laciniata</i> L.	SCLA1	N		X	X
ASTERACEAE	<i>Senecio plattensis</i> Nutt.	SEPL1	Y	X	X	X
ASTERACEAE	<i>Senecio spartioides</i> T. & G.	SESP1	Y			X
ASTERACEAE	<i>Solidago mollis</i> Bart.	SOMO1	Y	X		X
ASTERACEAE	<i>Taraxacum officinale</i> Weber	TAOF1	N		X	X
ASTERACEAE	<i>Thelesperma megapotanicum</i> (Spreng.) O. Ktze.	THME1	Y		X	
ASTERACEAE	<i>Tragopogon dubius</i> Scop.	TRDU1	N	X	X	X
BORAGINACEAE	<i>Lithospermum incisum</i> Lehm.	LIIN1	Y		X	X
BORAGINACEAE	<i>Mertensia lanceolata</i> (Pursh.) A. DC.	MELA1	Y	X		
BORAGINACEAE	<i>Onosmodium molle</i> Michx. var. <i>occidentale</i> (Mack.) Johnst.	ONMO1	Y			X
BRASSICACEAE	<i>Alyssum alyssoides</i> (L.) L.	ALAL1	N	X		
BRASSICACEAE	<i>Alyssum minus</i> (L.) Rothmaler var. <i>micranthus</i> (C. A. Mey.) Dudley	ALMI1	N	X	X	X
BRASSICACEAE	<i>Arabis hirsuta</i> (L.) Scop. var. <i>pynocarpa</i> (Hopkins) Rollins	ARHI1	Y	X		
BRASSICACEAE	<i>Camelina microcarpa</i> Andr. ex DC.	CAMI1	N	X	X	X
BRASSICACEAE	<i>Descurainia pinnata</i> (Walt.) Britt.	DEPI1	Y		X	X
BRASSICACEAE	<i>Descurainia richardsonii</i> (Sweet) Schultz	DERI1	Y		X	
BRASSICACEAE	<i>Lepidium densiflorum</i> Schrad.	LEDE1	Y			X
BRASSICACEAE	<i>Lesquerella montana</i> (A. Gray) Wats.	LEMO1	Y	X	X	X
BRASSICACEAE	<i>Sisymbrium altissimum</i> L.	SIAL1	N		X	
CACTACEAE	<i>Coryphantha missouriensis</i> (Sweet) Britt. & Rose	COM1	Y	X	X	X
CACTACEAE	<i>Echinocereus viridiflorus</i> Engelm.	ECV11	Y	X	X	X
CACTACEAE	<i>Pediocactus simpsonii</i> (Engelm.) Britt. & Rose	PES11	Y		X	
CARYOPHYLLACEAE	<i>Arenaria fendleri</i> A. Gray	ARFE2	Y	X	X	X
CARYOPHYLLACEAE	<i>Paronychia jamesii</i> T. & G.	PAJA1	Y	X	X	X
CARYOPHYLLACEAE	<i>Silene antirrhina</i> L.	SIAN1	Y	X	X	X
CARYOPHYLLACEAE	<i>Silene drummondii</i> Hook.	SIDR1	Y	X		
CHENOPodiaceae	<i>Chenopodium leptophyllum</i> Nutt. ex Moq.	CHLE2	Y		X	X
CLUSIACEAE	<i>Hypericum perforatum</i> L.	HYPE1	N	X	X	X
COMMELINACEAE	<i>Tradescantia occidentalis</i> (Britt.) Smyth	TROC1	Y	X	X	
CONVOLVULACEAE	<i>Convolvulus arvensis</i> L.	COAR1	N		X	
CONVOLVULACEAE	<i>Evolvulus nuttallianus</i> R. & S.	EVNU1	Y			X
CRASSULACEAE	<i>Sedum lanceolatum</i> Torr.	SELA1	Y	X		
CYPERACEAE	<i>Carex eleocharis</i> Bailey	CAEL1	Y			X
CYPERACEAE	<i>Carex filifolia</i> Nutt.	CAFI1	Y			X
CYPERACEAE	<i>Carex heliophila</i> Mack.	CAHE1	Y	X	X	X
EUPHORBIACEAE	<i>Euphorbia robusta</i> (Engelm.) Small	EIRO1	Y			X
FABACEAE	<i>Astragalus agrestis</i> Dougl. ex G. Don	ASAG1	Y	X	X	
FABACEAE	<i>Astragalus flexuosus</i> (Hook.) G. Don	ASFL1	Y		X	

Table 5-1. (cont.)

Family	Scientific Name	Speccode	Native	TR01	TR06	TR12
FABACEAE	<i>Astragalus shortianus</i> Nutt. ex T.&G.	ASSH1	Y	X	X	X
FABACEAE	<i>Dalea purpurea</i> Vent.	DAPU1	Y	X	X	X
FABACEAE	<i>Oxytropis lambertii</i> Pursh.	OXLA1	Y	X	X	X
FABACEAE	<i>Psoralea tenuiflora</i> Pursh.	PSTE1	Y	X	X	X
GERANIACEAE	<i>Erodium cicutarium</i> (L.) L'Her.	ERCI1	N			X
HYDROPHYLACEAE	<i>Phacelia heterophylla</i> Pursh.	PHHE1	Y		X	X
JUNCACEAE	<i>Juncus interior</i> Wieg.	JUIN1	Y	X		X
LILIACEAE	<i>Allium textile</i> A. Nels. & Macbr.	ALTE1	Y	X	X	X
LILIACEAE	<i>Leucocrinum montanum</i> Nutt.	LEMO2	Y		X	X
LINACEAE	<i>Linum perenne</i> L. var. <i>lewisii</i> (Pursh.) Eat. & Wright	LIPE1	Y		X	
NYCTAGINACEAE	<i>Mirabilis linearis</i> (Pursh.) Heimerl	MILI1	Y	X	X	X
ONAGRACEAE	<i>Calylophus serrulatus</i> (Nutt.) Raven	CASE2	Y	X		X
ONAGRACEAE	<i>Gaura coccinea</i> Pursh.	GACO1	Y		X	
OROBANCHACEAE	<i>Orobanche fasciculata</i> Nutt.	ORFA1	Y	X		X
POACEAE	<i>Agropyron smithii</i> Rydb.	AGSM1	Y	X	X	
POACEAE	<i>Andropogon gerardii</i> Vitman	ANGE1	Y	X	X	X
POACEAE	<i>Andropogon scoparius</i> Michx.	ANSC1	Y	X	X	X
POACEAE	<i>Aristida purpurea</i> Nutt. var. <i>robusta</i> (Merrill) A. Holmgren & N. Holmgr	ARLO1	Y	X	X	X
POACEAE	<i>Bouteloua curtipendula</i> (Michx.) Torr.	BOCU1	Y	X	X	X
POACEAE	<i>Bouteloua gracilis</i> (H. B. K.) Lag ex Griffiths	BOGR1	Y	X	X	X
POACEAE	<i>Bouteloua hirsuta</i> Lag	BOHI1	Y	X	X	X
POACEAE	<i>Bromus japonicus</i> Thunb. ex Murr.	BRJA1	N	X	X	X
POACEAE	<i>Bromus tectorum</i> L.	BRTE1	N	X	X	X
POACEAE	<i>Buchloe dactyloides</i> (Nutt.) Engelm.	BUDA1	Y	X	X	X
POACEAE	<i>Koeleria pyramidata</i> (Lam.) Beauv.	KOPY1	Y	X	X	X
POACEAE	<i>Muhlenbergia montana</i> (Nutt.) Hitchc.	MUMO1	Y	X		X
POACEAE	<i>Poa compressa</i> L.	POCO1	N	X	X	X
POACEAE	<i>Poa pratensis</i> L.	POPR1	N	X	X	X
POACEAE	<i>Sitanion hystris</i> (Nutt.) Sm. var. <i>brevifolium</i> (Sm.) Hitchc.	SIHY1	Y	X	X	X
POACEAE	<i>Sorghastrum nutans</i> (L.) Nash	SONU1	Y	X		X
POACEAE	<i>Sporobolus cryptandrus</i> (Torr.) A. Gray	SPCR1	Y	X	X	X
POACEAE	<i>Sporobolus heterolepis</i> (A. Gray) A. Gray	SPHE1	Y	X		
POACEAE	<i>Stipa comata</i> Trin. & Rupr.	STCO1	Y	X	X	X
POACEAE	<i>Stipa neomexicana</i> (Thur.) Scribn.	STNE1	Y		X	
POLEMONIACEAE	<i>Ipomopsis spicata</i> (Nutt.) V. Grant ssp. <i>spicata</i>	IPSP1	Y	X		
POLYGONACEAE	<i>Eriogonum alatum</i> Torr.	ERAL1	Y	X	X	X
PORTULACACEAE	<i>Talinum parviflorum</i> Nutt.	TAPA1	Y	X		
RANUNCULACEAE	<i>Delphinium nuttalianum</i> Pritz. ex Walpers	DENU1	Y			X
ROSACEAE	<i>Potentilla fissa</i> Nutt.	POFI1	Y	X		X
ROSACEAE	<i>Potentilla hippiana</i> Lehm.	POHI1	Y	X		
SANTALACEAE	<i>Comandra umbellata</i> (L.) Nutt.	COUM1	Y	X	X	X
SCROPHULARIACEAE	<i>Castilleja sessiliflora</i> Pursh.	CASE3	Y	X		X
SCROPHULARIACEAE	<i>Linaria dalmatica</i> (L.) Mill.	LIDA1	N		X	X
SCROPHULARIACEAE	<i>Penstemon secundiflorus</i> Benth.	PESE1	Y		X	
SCROPHULARIACEAE	<i>Penstemon virens</i> Penn.	PEV1	Y	X		X
SCROPHULARIACEAE	<i>Verbascum thapsus</i> L.	VETH1	N		X	
SOLANACEAE	<i>Solanum triflorum</i> Nutt.	SOTR1	Y			X
VIOLACEAE	<i>Viola nuttallii</i> Pursh.	VINU1	Y	X	X	X
BRASSICACEAE	<i>Draba reptans</i> (Lam.) Fern.	DRRE1	Y		X	X
ASTERACEAE	<i>Helianthus rigidus</i> (Cass.) Desf. ssp. <i>subrhomboideus</i> (Rydb.) Heiser	HERI1	Y	X		
BRASSICACEAE	<i>Erysimum capitatum</i> (Nutt.) DC.	ERCA2	Y	X	X	X
CACTACEAE	<i>Opuntia macrorhiza</i> Engelm.	OPMA1	Y	X	X	X
FABACEAE	<i>Astragalus tridactylus</i> Gray	ASTR1	Y		X	
ONAGRACEAE	<i>Oenothera howardii</i> (A. Nels.) W. L. Wagner	OEOH1	Y		X	
ASTERACEAE	<i>Solidago nana</i> Nutt.	SONA1	Y			X

Species Richness Summary

	Entire Community	TR01	TR06	TR12
Total # of species	122	81	83	84
Percent native species	84	86	81	83
Total # of families	35	26	23	28
Total # of monocots	28	23	24	21
Total # of dicots	94	58	59	63
Ratio monocots/dicots	0.3	0.4	0.41	0.33
Total # of forbs	92	56	58	62
Total # of graminoids	24	21	20	19
Total # of cactus	4	3	4	3
Total # of shrubs	2	1	1	0

Table 5-2. 1993–1998 xeric mixed grassland vegetation measurement comparisons

Data	TR01				TR06				TR12			
	1993	1994	1995	1998	1993	1994	1995	1998	1993	1994	1995	1998
# Species	76	88	90	81	68	89	98	83	68	91	83	84
% Native species	87	84	86	86	72	80	80	81	81	84	81	83
Total foliar cover	71.4	81.2	84.6	81.2	76.4	89.4	92.4	85.4	80.2	90.4	89.2	82.8
Total native cover	91.3	88.2	86.8	84.2	79.8	73.8	62.6	65.6	92.8	87.8	76.2	86.5
Cool-season graminoid cover	24.1	27.3	30.3	30.5	78.5	80.5	69.5	79.5	51.6	55.1	49.6	62.6
Warm-season graminoid cover	40.9	37.7	30.7	41.1	5.5	5.1	5.8	5.2	27.9	21.2	20.4	27.8
Forb cover	35	35	39	28.3	15.7	14.3	24.7	15	20.4	23	30	9.7
Basal vegetation cover	19.2	15.8	7.8		21.6	16.2	7.2			17.2	15	7.2
Litter cover	57	51.2	68.6		73.4	74.2	88			65.4	57.4	72.8
Rock cover	21.6	23.8	19.8		3.4	5.4	3.8			16.8	19.8	15.6
Bare ground cover	2.2	9.2	3.8		1.6	4.2	1			0.6	7.8	4.4

Note: Cover values are mean percentages (n = 5).

Table 5-3. 1998 xeric mixed grassland foliar cover summary

Family	Scientific Name	Speccode	Native	TR01			TR06			TR12		
				Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover
APIACEAE	<i>Lomatium orientale</i> Coulte & Rose	LOOR1	Y							20	0.20	0.24
ASTERACEAE	<i>Ambrosia psilostachya</i> DC.	AMPS1	Y	20	0.20	0.25	20	1.00	1.17	40	0.40	0.48
ASTERACEAE	<i>Antennaria parvifolia</i> Nutt.	ANPA1	Y	20	0.20	0.25						
ASTERACEAE	<i>Artemisia frigida</i> Willd.	ARFR1	Y	20	0.20	0.25	20	0.20	0.23			
ASTERACEAE	<i>Artemisia ludoviciana</i> Nutt. var. <i>ludoviciana</i>	ARLU1	Y							40	0.80	0.97
ASTERACEAE	<i>Aster porteri</i> Gray	ASPO1	Y	100	11.80	14.53				80	1.20	1.45
ASTERACEAE	<i>Carduus nutans</i> L. ssp. <i>macrolepis</i> (Peterm.) Kazmi	CANU1	N				20	0.20	0.23			
ASTERACEAE	<i>Centaurea diffusa</i> Lam.	CEDI1	N	20	0.20	0.25						
ASTERACEAE	<i>Chrysopsis villosa</i> Pursh.	CHVI1	Y	40	0.40	0.49						
ASTERACEAE	<i>Helianthus pumilus</i> Nutt.	HEPU1	Y				20	0.20	0.23			
ASTERACEAE	<i>Lactuca serriola</i> L.	LASE1	N							20	0.20	0.24
ASTERACEAE	<i>Liatris punctata</i> Hook.	LIPU1	Y	80	2.40	2.96				80	2.20	2.66
ASTERACEAE	<i>Tragopogon dubius</i> Scop.	TRDU1	N				20	0.20	0.23			
BORAGINACEAE	<i>Lithospermum incisum</i> Lehm.	LIIN1	Y							20	0.20	0.24
BRASSICACEAE	<i>Alyssum minus</i> (L.) Rothmaler var. <i>micranthus</i> (C. A. Mey.) Dudley	ALMI1	N	20	0.20	0.25	40	1.40	1.64	60	0.80	0.97
BRASSICACEAE	<i>Descurainia pinnata</i> (Walt.) Britt.	DEPI1	Y							20	0.20	0.24
BRASSICACEAE	<i>Lesquerella montana</i> (A. Gray) Wats.	LEMO1	Y	20	0.20	0.25						
CARYOPHYLLACEAE	<i>Arenaria fendleri</i> A. Gray	ARFE2	Y	100	4.00	4.93				80	1.00	1.21
CARYOPHYLLACEAE	<i>Paronychia jamesii</i> T. & G.	PAJA1	Y	40	0.40	0.49						
CYPERACEAE	<i>Carex filifolia</i> Nutt.	CAFI1	Y				20	0.20	0.23			
CYPERACEAE	<i>Carex heliophila</i> Mack.	CAHE1	Y	80	5.80	7.14	60	1.60	1.87	100	4.00	4.83
EUPHORBIACEAE	<i>Euphorbia robusta</i> (Engelm.) Small	EURO1	Y							40	0.40	0.48
FABACEAE	<i>Dalea purpurea</i> Vent	DAPU1	Y	20	0.20	0.25						
FABACEAE	<i>Oxytropis lambertii</i> Pursh.	OXL1	Y							20	0.20	0.24
FABACEAE	<i>Psoralea tenuiflora</i> Pursh.	PSTE1	Y	80	1.40	1.72				20	0.20	0.24
LINACEAE	<i>Linum perenne</i> L. var. <i>lewisii</i> (Pursh.) Eat. & Wright	LIPE1	Y				40	0.40	0.47			
ONAGRACEAE	<i>Calylophus serrulatus</i> (Nutt.) Raven	CASE2	Y	20	0.20	0.25						
POACEAE	<i>Andropogon gerardii</i> Vitman	ANGE1	Y	100	9.60	11.82	40	0.80	0.94	100	12.20	14.73
POACEAE	<i>Andropogon scoparius</i> Michx.	ANSC1	Y	80	1.40	1.72	40	0.40	0.47			
POACEAE	<i>Aristida purpurea</i> Nutt. var. <i>robusta</i> (Merrill) A. Holmgren & N. Holmgr	ARLO1	Y	20	0.20	0.25	60	0.60	0.70			
POACEAE	<i>Bouteloua curtipendula</i> (Michx.) Torr.	BOCU1	Y	100	3.00	3.69	80	1.60	1.87	100	3.60	4.35
POACEAE	<i>Bouteloua gracilis</i> (H. B. K.) Lag ex Griffiths	BOGR1	Y	80	1.60	1.97	80	1.60	1.87	100	3.00	3.62
POACEAE	<i>Bouteloua hirsuta</i> Lag	BOHI1	Y	60	1.00	1.23				40	0.40	0.48
POACEAE	<i>Bromus japonicus</i> Thunb. ex Murr.	BRJA1	N	40	0.40	0.49	100	7.40	8.67	80	1.80	2.17
POACEAE	<i>Bromus tectorum</i> L.	BRTE1	N	20	0.40	0.49				20	0.80	0.97
POACEAE	<i>Koeleria pyramidata</i> (Lam.) Beauv.	KOPY1	Y	80	1.60	1.97				80	1.20	1.45
POACEAE	<i>Muhlenbergia montana</i> (Nutt.) Hitchc.	MUMO1	Y	80	14.80	18.23				100	2.20	2.66
POACEAE	<i>Poa compressa</i> L.	POCO1	N	80	6.60	8.13	60	4.20	4.92	100	6.20	7.49
POACEAE	<i>Poa pratensis</i> L.	POPR1	N	20	5.00	6.16	80	6.80	7.96	60	1.40	1.69
POACEAE	<i>Sitanion hystricoides</i> (Nutt.) Sm. var. <i>brevifolium</i> (Sm.) Hitchc.	SIHY1	Y	40	0.80	0.99	40	0.40	0.47	20	0.20	0.24

Table 5-3. (cont.)

Family	Scientific Name	Speccode	Native	TR01			TR06			TR12		
				Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover	Frequency	Absolute Cover	Relative Cover
POACEAE	<i>Sorghastrum nutans</i> (L.) Nash	SONU1	Y	80	1.20	1.48				40	1.60	1.93
POACEAE	<i>Sporobolus heterolepis</i> (A. Gray) A. Gray	SPHE1	Y	60	0.80	0.99						
POACEAE	<i>Stipa comata</i> Trin. & Rupr.	STCO1	Y	100	4.00	4.93	100	43.40	50.82	100	36.20	43.72
POACEAE	<i>Stipa neomexicana</i> (Thur.) Scribn.	STNE1	Y				20	3.60	4.22			
POLYGONACEAE	<i>Eriogonum alatum</i> Torr.	ERAL1	Y	40	0.40	0.49						
PORTULACACEAE	<i>Talinum parviflorum</i> Nutt.	TAPA1	Y	20	0.20	0.25						
SANTALACEAE	<i>Comandra umbellata</i> (L.) Nutt.	COUM1	Y	40	0.40	0.49						
SCROPHULARIACEAE	<i>Linaria dalmatica</i> (L.) Mill.	LIDA1	N				100	9.20	10.77			
	Total foliar cover				81.20	100.00		85.40	100.00		82.80	100.00
	Total native foliar cover				68.40	84.24		56.00	65.57		71.60	86.47

Note: Absolute cover = Absolute foliar cover is the percentage of the number of hits on a species out of the total number of hits possible at a site (500).

Relative cover = Relative foliar cover is the number of hits on a species relative to the total number of all vegetative hits recorded per site (i.e., the percent of vegetative cover the species represented).

All cover values presented are means (n = 5).

Table 5-4. 1998 xeric mixed grassland site plant frequencies

Family	Scientific Name	Native	Speccode	TR01		TR06		TR12	
				Spring	Summer	Spring	Summer	Spring	Summer
AGAVACEAE	<i>Yucca glauca</i> Nutt.	Y	YUGL1			20	12		
APIACEAE	<i>Lomatium orientale</i> Coulte & Rose	Y	LOOR1	92	28	72	4	100	48
ASCLEPIADACEAE	<i>Asclepias viridiflora</i> Raf.	Y	ASVI1	4	4			4	4
ASTERACEAE	<i>Achillea millefolium</i> L. ssp. <i>lanulosa</i> (Nutt.) Piper	Y	ACMI1	4	4				
ASTERACEAE	<i>Ambrosia psilostachya</i> DC.	Y	AMPS1	28	16	12	12	24	28
ASTERACEAE	<i>Antennaria parvifolia</i> Nutt.	Y	ANPA1	4	4			4	4
ASTERACEAE	<i>Artemisia frigida</i> Willd.	Y	ARFR1	16	16	20	20	8	4
ASTERACEAE	<i>Artemisia ludoviciana</i> Nutt. var. <i>ludoviciana</i>	Y	ARLU1			8		16	12
ASTERACEAE	<i>Aster falcatus</i> Lindl.	Y	ASFA1	4	4				
ASTERACEAE	<i>Aster porteri</i> Gray	Y	ASPO1	88	88			40	44
ASTERACEAE	<i>Carduus nutans</i> L. ssp. <i>macrolepis</i> (Peterm.) Kazmi	N	CANU1			24	16		
ASTERACEAE	<i>Centaurea diffusa</i> Lam.	N	CEDI1	16	4			4	8
ASTERACEAE	<i>Chrysopsis fulcrata</i> Greene	Y	CHFU1	8	8				
ASTERACEAE	<i>Chrysopsis villosa</i> Pursh.	Y	CHVI1	64	52			28	32
ASTERACEAE	<i>Cirsium arvense</i> (L.) Scop.	N	CIAR1			8	8		
ASTERACEAE	<i>Cirsium undulatum</i> (Nutt.) Spreng.	Y	CIUN1			8	4		
ASTERACEAE	<i>Erigeron flagellaris</i> A. Gray	Y	ERFL1					20	
ASTERACEAE	<i>Gaillardia aristata</i> Pursh.	Y	GAAR1	28	16			12	4
ASTERACEAE	<i>Helianthus pumilus</i> Nutt.	Y	HEPU1	4	4	4	4		
ASTERACEAE	<i>Lactuca serriola</i> L.	N	LASE1			52	28		
ASTERACEAE	<i>Liatris punctata</i> Hook.	Y	LIPU1	76	72		4	68	76
ASTERACEAE	<i>Microseris cuspidata</i> (Pursh.) Sch. Bip.	Y	MICU1	8				12	
ASTERACEAE	<i>Scorzonera laciniata</i> L.	N	SCLA1			8			4
ASTERACEAE	<i>Senecio plattensis</i> Nutt.	Y	SEPL1	40	20				
ASTERACEAE	<i>Senecio spartioides</i> T. & G.	Y	SESP1					8	8
ASTERACEAE	<i>Solidago mollis</i> Bart.	Y	SOMO1	4	4				
ASTERACEAE	<i>Taraxacum officinale</i> Weber	N	TAOF1			12	4	8	
ASTERACEAE	<i>Tragopogon dubius</i> Scop.	N	TRDU1	20	20	68	40	8	12
BORAGINACEAE	<i>Lithospermum incisum</i> Lehm.	Y	LIIN1				12		4
BORAGINACEAE	<i>Mertensia lanceolata</i> (Pursh.) A. DC.	Y	MELA1	4					
BRASSICACEAE	<i>Alyssum alyssoides</i> (L.) L.	N	ALAL1	12	4				
BRASSICACEAE	<i>Alyssum minus</i> (L.) Rothmaler var. <i>micranthus</i> (C. A. Mey.) Dudley	N	ALMI1	20	16	36	40	80	72
BRASSICACEAE	<i>Camelina microcarpa</i> Andrz. ex DC.	N	CAMI1		4	76	24	44	24
BRASSICACEAE	<i>Descurainia pinnata</i> (Walt.) Britt.	Y	DEPI1			16			
BRASSICACEAE	<i>Draba reptans</i> (Lam.) Fern.	Y	DRRE1	16		8		28	12
BRASSICACEAE	<i>Erysimum capitatum</i> (Nutt.) DC.	Y	ERCA2	40	20	12	4		4
BRASSICACEAE	<i>Lepidium densiflorum</i> Schrad.	Y	LEDE1					4	4
BRASSICACEAE	<i>Lesquerella montana</i> (A. Gray) Wats.	Y	LEMO1	68	56	32	20	44	36
BRASSICACEAE	<i>Sisymbrium altissimum</i> L.	N	SIAL1			4			
CACTACEAE	<i>Echinocereus viridiflorus</i> Engelm.	Y	ECV1	40	52		4	60	52
CACTACEAE	<i>Opuntia macrorhiza</i> Engelm.	Y	OPMA1	28	20	12	12	32	36

Table 5-4. (cont.)

Family	Scientific Name	Native	Speccode	TR01		TR06		TR12	
				Spring	Summer	Spring	Summer	Spring	Summer
CARYOPHYLLACEAE	<i>Arenaria fendleri</i> A. Gray	Y	ARFE2	76	80		4	72	72
CARYOPHYLLACEAE	<i>Paronychia jamesii</i> T. & G.	Y	PAJA1	48	52			8	8
CARYOPHYLLACEAE	<i>Silene antirrhina</i> L.	Y	SIAN1	12					
CARYOPHYLLACEAE	<i>Silene drummondii</i> Hook.	Y	SIDR1	4	4			4	4
CLUSIACEAE	<i>Hypericum perforatum</i> L.	N	HYPE1	4	40				4
COMMELINACEAE	<i>Tradescantia occidentalis</i> (Britt.) Smyth	Y	TROC1	4		8			
CYPERACEAE	<i>Carex filifolia</i> Nutt.	Y	CAFI1			8	4		
CYPERACEAE	<i>Carex heliophila</i> Mack.	Y	CAHE1	84	92	28	32	100	100
FABACEAE	<i>Astragalus agrestis</i> Dougl. ex G. Don	Y	ASAG1	4	4				
FABACEAE	<i>Astragalus shortianus</i> Nutt. ex T.&G.	Y	ASSH1			4			
FABACEAE	<i>Astragalus tridactylicus</i> Gray	Y	ASTR1					4	
FABACEAE	<i>Dalea purpurea</i> Vent	Y	DAPU1	16	16	4	4	4	
FABACEAE	<i>Psoralea tenuiflora</i> Pursh.	Y	PSTE1	60	56	12	8	48	44
JUNCACEAE	<i>Juncus interior</i> Wieg.	Y	JUIN1						4
LILIACEAE	<i>Allium textile</i> A. Neils. & Macbr.	Y	ALTE1	20		12		28	20
LILIACEAE	<i>Leucocrinum montanum</i> Nutt.	Y	LEMO2			4			
LINACEAE	<i>Linum perenne</i> L. var. <i>lewisii</i> (Pursh.) Eat. & Wright	Y	LIPE1			48	32		
NYCTAGINACEAE	<i>Mirabilis linearis</i> (Pursh.) Heimerl	Y	MILI1		4				12
ONAGRACEAE	<i>Gaura coccinea</i> Pursh.	Y	GACO1			4			
OROBANCHACEAE	<i>Orobanche fasciculata</i> Nutt.	Y	ORFA1	4				32	
POACEAE	<i>Agropyron smithii</i> Rydb.	Y	AGSM1		4	4			
POACEAE	<i>Andropogon gerardii</i> Vitman	Y	ANGE1	68	76	8	4	60	60
POACEAE	<i>Andropogon scoparius</i> Michx.	Y	ANSC1	60	68	4	4		4
POACEAE	<i>Aristida purpurea</i> Nutt. var. <i>robusta</i> (Merrill) A. Holmgren & N. Holmgr	Y	ARLO1		4	8	12		16
POACEAE	<i>Bouteloua curtipendula</i> (Michx.) Torr.	Y	BOCU1	52	60	60	72	60	84
POACEAE	<i>Bouteloua gracilis</i> (H. B. K.) Lag ex Griffiths	Y	BOGR1	52	68	44	72	64	84
POACEAE	<i>Bouteloua hirsuta</i> Lag	Y	BOHI1	40	68		12	36	28
POACEAE	<i>Bromus japonicus</i> Thunb. ex Murr.	N	BRJA1	20	8	68	60	40	52
POACEAE	<i>Bromus tectorum</i> L.	N	BRTE1	8	4			4	8
POACEAE	<i>Buchloe dactyloides</i> (Nutt.) Engelm.	Y	BUDA1	8				16	24
POACEAE	<i>Koeleria pyramidata</i> (Lam.) Beauv.	Y	KOPY1	76	76	16	12	56	48
POACEAE	<i>Muhlenbergia montana</i> (Nutt.) Hitchc.	Y	MUMO1	60	64			4	8
POACEAE	<i>Poa compressa</i> L.	N	POCO1	36	36	20	16	36	44
POACEAE	<i>Poa pratensis</i> L.	N	POPR1	28	24	32	28	20	16
POACEAE	<i>Sitanion hystrix</i> (Nutt.) Sm. var. <i>brevifolium</i> (Sm.) Hitchc.	Y	SIHY1	48	44	8	4		
POACEAE	<i>Sorghastrum nutans</i> (L.) Nash	Y	SONU1	4	12			4	8
POACEAE	<i>Sporobolus heterolepis</i> (A. Gray) A. Gray	Y	SPHE1	16	16				
POACEAE	<i>Stipa comata</i> Trin. & Rupr.	Y	STCO1	24	32	88	92	100	100
POACEAE	<i>Stipa neomexicana</i> (Thur.) Scribn.	Y	STNE1			20	20		
POLYGONACEAE	<i>Eriogonum alatum</i> Torr.	Y	ERAL1	68	68	4		4	4
PORTULACACEAE	<i>Talinum parviflorum</i> Nutt.	Y	TAPA1	24	32				

Table 5-4. (cont.)

Family	Scientific Name	Native	Speccode	TR01		TR06		TR12	
				Spring	Summer	Spring	Summer	Spring	Summer
ROSACEAE	Potentilla fissa Nutt.	Y	POFI1	4	4				
ROSACEAE	Potentilla hippiana Lehm.	Y	POHI1	8	8				
SANTALACEAE	Comandra umbellata (L.) Nutt.	Y	COUM1	28	28				
SCROPHULARIACEAE	Castilleja sessiliflora Pursh.	Y	CASE3	12	4				
SCROPHULARIACEAE	Linaria dalmatica (L.) Mill.	N	LIDA1			100	100		12
SCROPHULARIACEAE	Penstemon secundiflorus Benth.	Y	PESE1			4			
SCROPHULARIACEAE	Penstemon virens Penn.	Y	PEVI1	8					
SOLANACEAE	Solanum triflorum Nutt.	Y	SOTR1						4
VIOLACEAE	Viola nuttallii Pursh.	Y	VINU1	16		48		56	

Note: All values are percentages based on n = 25.

Table 5-5. Plant species with large or consistent increases or declines in relative foliar cover (1993–1998)

Station/Scientific Name	Speccode	1993	1994	1995	1998	Change
TR01						
Andropogon scoparius	ANSC1	11.76	10.34	5.44	1.72	-10.04
Muhlenbergia montana	MUMO1	9.80	8.37	8.75	18.23	8.42
Aster porteri	ASPO1	3.92	10.34	25.06	14.53	10.61
TR06						
Bromus japonicus	BRJA1	0.79	2.46	6.06	8.67	7.88
TR12						
Andropogon scoparius	ANSC1	5.49	2.88	0.90	0.00	-5.49
Arenaria fendleri	ARFE2	5.24	6.19	2.47	1.21	-4.03
Psorelea tenuiflora	PSTE1	4.24	2.43	1.57	0.24	-4.00

Note: Cover values are all mean percentages (n = 5).

Appendix B

Rocky Flats Plant List

Plant Species Known to Occur at Rocky Flats Environmental Technology Site, May 1999

Family	Scientific Name	Speccode	Common Name
ACERACEAE	Acer glabrum Torr.	ACGL1	Mountain Maple
ACERACEAE	Acer negundo L. var. interius (Britt.) Sarg.	ACNE1	Box-elder
AGAVACEAE	Yucca glauca Nutt.	YUGL1	Yucca
ALISMATACEAE	Alisma trivale Pursh	ALTR1	American Water Plantain
ALISMATACEAE	Sagittaria latifolia Willd.	SALA1	Common Arrowhead
AMARANTHACEAE	Amaranthus albus L.	AMAL2	Tumbleweed
AMARANTHACEAE	Amaranthus retroflexus L.	AMRE1	Rough Pigweed
ANACARDIACEAE	Rhus aromatica Ait. var. trilobata (Nutt.) A. Gray	RHAR1	Fragrant Sumac
ANACARDIACEAE	Toxicodendron rydbergii (Small) Greene	TORY1	Poison Ivy
APIACEAE	Cicuta maculata L. var. angustifolia Hook.	CIMA1	Water Hemlock
APIACEAE	Conium maculatum L.	COMA1	Poison Hemlock
APIACEAE	Daucus carota L.	DACA2	Wild Carrot
APIACEAE	Harbouria trachyleura (Gray) C. & R.	HATR1	Whiskbroom Parsley
APIACEAE	Heracleum sphondylium L. ssp. montanum (Schleich.) Briq.	HESP1	Cow Parsnip
APIACEAE	Ligusticum porteri C. & R.	LIPO1	Porter's Lovage
APIACEAE	Lomatium orientale Coulte. & Rose	LOOR1	Wild Parsley
APIACEAE	Musineon divaricatum (Pursh.) Nutt. var. hookeri T. & G.	MUDI1	Musineon
APIACEAE	Osmorhiza chilensis H. & A.	OSCH1	Sweet Cicely
APIACEAE	Osmorhiza longistylis (Torr.) DC var. longistylis	OSLO1	Anise Root
APOCYNACEAE	Apocynum androsaemifolium L.	APAN1	Spreading Dogbane
APOCYNACEAE	Apocynum cannabinum L.	APCA1	Hemp Dogbane
ASCLEPIADACEAE	Asclepias incarnata L.	ASIN1	Swamp Milkweed
ASCLEPIADACEAE	Asclepias pumila (Gray) Vail	ASPU1	Plains Milkweed
ASCLEPIADACEAE	Asclepias speciosa Torr.	ASSP1	Showy Milkweed
ASCLEPIADACEAE	Asclepias stenophylla A. Gray	ASST1	Narrow-leaved Milkweed
ASCLEPIADACEAE	Asclepias viridiflora Raf.	ASVI1	Green Milkweed
ASTERACEAE	Achillea millefolium L. ssp. lanulosa (Nutt.) Piper	ACMI1	Yarrow
ASTERACEAE	Agoseris glauca (Pursh.) Dietr.	AGGL1	False Dandelion
ASTERACEAE	Ambrosia artemisiifolia L.	AMAR1	Common Ragweed
ASTERACEAE	Ambrosia psilostachya DC.	AMPS1	Western Ragweed
ASTERACEAE	Ambrosia trifida L.	AMTR1	Giant Ragweed
ASTERACEAE	Antennaria microphylla Rydb.	ANMI1	Pink Pussytoes
ASTERACEAE	Antennaria parvifolia Nutt.	ANPA1	Pussytoes
ASTERACEAE	Anthemis cotula L.	ANCO1	Dog Fennel
ASTERACEAE	Arctium minus Bernh.	ARMI1	Burdock
ASTERACEAE	Arnica fulgens Pursh.	ARFU1	Arnica
ASTERACEAE	Artemisia campestris L. ssp. caudata (Michx.) Hall & Clem.	ARCA1	Western Sagewort
ASTERACEAE	Artemisia dracunculus L.	ARDR1	Silky Wormwood
ASTERACEAE	Artemisia frigida Willd.	ARFR1	Silver Sage
ASTERACEAE	Artemisia ludoviciana Nutt. var. ludoviciana	ARLU1	White Sage
ASTERACEAE	Aster falcatus Lindl.	ASFA1	Aster
ASTERACEAE	Aster fendleri A. Gray	ASFE1	Fendler's Aster
ASTERACEAE	Aster hesperius A. Gray var. hersperius	ASHE1	Panicled Aster
ASTERACEAE	Aster laevis L. var. geyeri A. Gray	ASLA1	Smooth Blue Aster
ASTERACEAE	Aster porteri Gray	ASPO1	Aster
ASTERACEAE	Bidens cernua L.	BICE1	Nodding Beggarticks
ASTERACEAE	Bidens frondosa L.	BIFR1	Beggar-ticks
ASTERACEAE	Carduus nutans L. ssp. macrolepis (Peterm.) Kazmi	CANU1	Musk Thistle
ASTERACEAE	Centaurea diffusa Lam.	CEDI1	Diffuse Knapweed
ASTERACEAE	Centaurea repens L.	CERE1	Russian Knapweed
ASTERACEAE	Chrysanthemum leucanthemum L.	CHLE1	Ox-eye Daisy
ASTERACEAE	Chrysopsis fulcrata Greene	CHFU1	Golden Aster
ASTERACEAE	Chrysopsis villosa Pursh.	CHV1	Golden Aster
ASTERACEAE	Chrysothamnus nauseosus (Pall.) Britt. ssp. graveolens (Nutt.) Piper	CHNA1	Greenplume Rabbitbrush
ASTERACEAE	Chrysothamnus nauseosus (Pall.) Britt. ssp. nauseosus	CHNA2	Rubber Rabbitbrush
ASTERACEAE	Cichorium intybus L.	CIIN1	Common Chicory
ASTERACEAE	Cirsium arvense (L.) Scop.	CIAR1	Canada Thistle
ASTERACEAE	Cirsium flodmanni (Rydb.) Arthur	CIFL1	Flodman's Thistle
ASTERACEAE	Cirsium ochrocentrum A. Gray	CIOC1	Yellow Spine Thistle
ASTERACEAE	Cirsium undulatum (Nutt.) Spreng.	CIUN1	Wavyleaf Thistle
ASTERACEAE	Cirsium vulgare (Savi) Ten.	CIVU1	Bull Thistle
ASTERACEAE	Conyza canadensis (L.) Cronq.	COCA1	Horseweed
ASTERACEAE	Crepis occidentalis Nutt.	CROC1	Hawksbeard
ASTERACEAE	Crepis runcinata (James) T. & G.	CRRU1	Hawksbeard
ASTERACEAE	Dyssodia papposa (Vent) Hitchc.	DYP1	Fetid Marigold

Plant Species Known to Occur at Rocky Flats Environmental Technology Site, May 1999 (cont.)

Family	Scientific Name	Speccode	Common Name
ASTERACEAE	<i>Erigeron canus</i> A. Gray	ERCA1	Fleabane
ASTERACEAE	<i>Erigeron compositus</i> Pursh var. <i>dicoideus</i> A. Gray	ERCO1	
ASTERACEAE	<i>Erigeron divergens</i> T. & G.	ERDI1	Fleabane
ASTERACEAE	<i>Erigeron flagellaris</i> A. Gray	ERFL1	Fleabane
ASTERACEAE	<i>Erigeron pumilus</i> Nutt.	ERPU1	Fleabane
ASTERACEAE	<i>Erigeron speciosa</i> (Lindl.) DC. var. <i>macranthus</i> (Nutt.) Cronq.	ERSP1	Oregon Fleabane
ASTERACEAE	<i>Erigeron strigosus</i> Muhl. ex Willd.	ERST1	Daisy Fleabane
ASTERACEAE	<i>Erigeron vetensis</i> Rydb.	ERVE1	LaVeta Fleabane
ASTERACEAE	<i>Gaillardia aristata</i> Pursh.	GAAR1	Blanket Flower
ASTERACEAE	<i>Gnaphalium chilense</i> Spreng.	GNCH1	Cotton-batting
ASTERACEAE	<i>Grindelia squarrosa</i> (Pursh.) Dun.	GRSQ1	Curly-top Gumweed
ASTERACEAE	<i>Gutierrezia sarothrae</i> (Pursh.) Britt. & Rusby	GUSA1	Snakeweed
ASTERACEAE	<i>Happlopappus spinulosus</i> (Pursh) DC.	HASP1	Cutleaf Ironplant
ASTERACEAE	<i>Helianthus annuus</i> L.	HEAN1	Common Sunflower
ASTERACEAE	<i>Helianthus ciliaris</i> DC.	HECI1	Texas Blue Weed
ASTERACEAE	<i>Helianthus maximilianii</i> Schrad.	HEMA1	Maximilian Sunflower
ASTERACEAE	<i>Helianthus nuttallii</i> T. & G.	HENU1	Nuttall's Sunflower
ASTERACEAE	<i>Helianthus petiolaris</i> Nutt.	HEPE1	Plains Sunflower
ASTERACEAE	<i>Helianthus pumilus</i> Nutt.	HEPU1	Sunflower
ASTERACEAE	<i>Helianthus rigidus</i> (Cass.) Desf. ssp. <i>subrhomboideus</i> (Rydb.) Heiser	HERI1	Stiff Sunflower
ASTERACEAE	<i>Helimeris multiflora</i> Nuttall	HEMU1	Showy Goldeneye
ASTERACEAE	<i>Hymenopappus filifolius</i> Hook. var. <i>cinereus</i> (Rydb.) I. M. Johnst.	HYFI1	Hymenopappus
ASTERACEAE	<i>Iva axillaris</i> Pursh.	IVAX1	Poverty Weed
ASTERACEAE	<i>Iva xanthifolia</i> Nutt.	IVXA1	Marsh Elder
ASTERACEAE	<i>Kuhnia chlorolepis</i> Woot. & Standl.	KUCH1	False Boneset
ASTERACEAE	<i>Kuhnia eupatorioides</i> L.	KUEU1	False Boneset
ASTERACEAE	<i>Lactuca oblongifolia</i> Nutt.	LAOB1	Blue Lettuce
ASTERACEAE	<i>Lactuca serriola</i> L.	LASE1	Prickly Lettuce
ASTERACEAE	<i>Leucelene ericoides</i> (Torr.) Greene	LEER1	White Aster
ASTERACEAE	<i>Liatris punctata</i> Hook.	LIPU1	Blazing Star
ASTERACEAE	<i>Machaeranthera bigelovii</i> (Gray) Greene	MABI1	Bigelovi's Tansy Aster
ASTERACEAE	<i>Machaeranthera canescens</i> (Pursh) A. Gray	MACA1	Hoary Aster
ASTERACEAE	<i>Microseris cuspidata</i> (Pursh.) Sch. Bip.	MICU1	False Dandelion
ASTERACEAE	<i>Onopordum acanthium</i> L.	ONAC1	Scotch Thistle
ASTERACEAE	<i>Picradeniopsis oppositifolia</i> (Nutt.) Rydb.	PIOP1	Picradeniopsis
ASTERACEAE	<i>Ratibida columnifera</i> (Nutt.) Woot. & Standl.	RACO1	Prairie Coneflower
ASTERACEAE	<i>Rudbeckia ampla</i> Nelson	RUAM1	Goldenglow
ASTERACEAE	<i>Scorzonera laciniata</i> L.	SCLA1	False Salsify
ASTERACEAE	<i>Senecio fendleri</i> Gray	SEFE1	Groundsel
ASTERACEAE	<i>Senecio integerrimus</i> Nutt.	SEIN1	Groundsel
ASTERACEAE	<i>Senecio plattensis</i> Nutt.	SEPL1	Prairie Ragwort
ASTERACEAE	<i>Senecio spartioides</i> T. & G.	SESP1	Groundsel
ASTERACEAE	<i>Senecio tridenticulatus</i> Rydb.	SETR1	Groundsel
ASTERACEAE	<i>Solidago canadensis</i> L.	SOC1	Canada Goldenrod
ASTERACEAE	<i>Solidago gigantea</i> Ait.	SOGI1	Late Goldenrod
ASTERACEAE	<i>Solidago missouriensis</i> Nutt.	SOMI1	Prairie Goldenrod
ASTERACEAE	<i>Solidago mollis</i> Bart.	SOMO1	Soft Goldenrod
ASTERACEAE	<i>Solidago nana</i> Nutt.	SONA1	Low Goldenrod
ASTERACEAE	<i>Solidago rigida</i> L.	SORI1	Rigid Goldenrod
ASTERACEAE	<i>Sonchus arvensis</i> L. ssp. <i>uglinosus</i> (Bieb.) Nyman	SOAR2	Field Sow Thistle
ASTERACEAE	<i>Sonchus asper</i> (L.) Hill	SOAS1	Prickly Sow Thistle
ASTERACEAE	<i>Stephanomeria pauciflora</i> (Torr.) A. Nels.	STPA1	Wire Lettuce
ASTERACEAE	<i>Taraxacum laevigatum</i> (Willd.) DC.	TALA1	Red Seeded Dandelion
ASTERACEAE	<i>Taraxacum officinale</i> Weber	TAOF1	Dandelion
ASTERACEAE	<i>Thelesperma megapotanicum</i> (Spreng.) O. Ktze.	THME1	Greenthread
ASTERACEAE	<i>Townsendia grandiflora</i> (Nutt.)	TOGR1	Easter Daisy
ASTERACEAE	<i>Townsendia hookeri</i> Beaman	TOHO1	Easter Daisy
ASTERACEAE	<i>Tragopogon dubius</i> Scop.	TRDU1	Goat's Beard
ASTERACEAE	<i>Tragopogon porrifolius</i> L.	TRPO1	Salsify
ASTERACEAE	<i>Xanthium strumarium</i> L.	XAST1	Cocklebur
BERBERIDACEAE	<i>Berberis repens</i> Lindl.	BERE1	Oregon Grape
BETULACEAE	<i>Alnus incana</i> (L.) Moench ssp. <i>tenuifolia</i> (Nuttall) Breitung	ALIN1	Alder
BETULACEAE	<i>Betula occidentalis</i> Hook.	BEOC1	Water Birch
BORAGINACEAE	<i>Asperugo procumbens</i> L.	ASPR1	Madwort
BORAGINACEAE	<i>Cryptantha virgata</i> (Porter) Payson	CRVI1	Miners Candle

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BORAGINACEAE	Cynoglossum officinale L.	CYOF1	Hound's Tongue
BORAGINACEAE	Hackelia floribunda (Lehm.) I. M. Johnst.	HAFL1	Large-flowered Stickseed
BORAGINACEAE	Lappula redowskii (Hornem.) Greene	LARE1	Stickseed
BORAGINACEAE	Lithospermum incisum Lehm.	LIIN1	Puccoon
BORAGINACEAE	Lithospermum multiflorum Torr.	LIMU1	
BORAGINACEAE	Mertensia lanceolata (Pursh.) A. DC.	MELA1	Bluebells
BORAGINACEAE	Onosmodium molle Michx. var. occidentale (Mack.) Johnst.	ONMO1	False Gromwell
BORAGINACEAE	Plagiobothrys scouleri (H. & A.) I. M. Johnst.	PLSC1	Popcorn Flower
BRASSICACEAE	Alyssum alyssoides (L.) L.	ALAL1	Pale Alyssum
BRASSICACEAE	Alyssum minus (L.) Rothmaler var. micranthus (C. A. Mey.) Dudley	ALMI1	Alyssum
BRASSICACEAE	Arabis fendleri (S. Wats.) Greene var. fendleri	ARFE3	Rock Cress
BRASSICACEAE	Arabis glabra (L.) Bernh.	ARGL1	Tower Mustard
BRASSICACEAE	Arabis hirsuta (L.) Scop. var. pynocarpa (Hopkins) Rollins	ARHI1	Rock Cress
BRASSICACEAE	Barbara vulgaris R. Br.	BAVU1	Yellowrocket Wintercress
BRASSICACEAE	Camelina microcarpa Andrz. ex DC.	CAMI1	Small-seeded False Flax
BRASSICACEAE	Capsella bursa-pastoris (L.) Medic.	CABU1	Shepherd's Purse
BRASSICACEAE	Cardaria chalepensis (L.) Hand-Mazz	CACH1	Lens-padded Hoary Cress
BRASSICACEAE	Cardaria draba (L.) Desv.	CADR1	Hoary Cress
BRASSICACEAE	Chorispora tenella (Pall.) DC.	CHTE1	Blue Mustard
BRASSICACEAE	Conringia orientalis (L.) Dum.	COOR1	Hare's-ear Mustard
BRASSICACEAE	Descurainia pinnata (Walt.) Britt.	DEPI1	Tansy Mustard
BRASSICACEAE	Descurainia richardsonii (Sweet) Schultz	DERI1	Tansy Mustard
BRASSICACEAE	Descurainia sophia (L.) Webb ex Prantl.	DESO1	Flixweed
BRASSICACEAE	Draba nemorosa L.	DRNE1	Yellow Whitlowort
BRASSICACEAE	Draba reptans (Lam.) Fern.	DRRE1	White Whitlowort
BRASSICACEAE	Erysimum capitatum (Nutt.) DC.	ERCA2	Western Wallflower
BRASSICACEAE	Erysimum repandum L.	ERRE1	Bushy Wallflower
BRASSICACEAE	Hesperis matronalis L.	HEMA2	Dame's Rocket
BRASSICACEAE	Lepidium campestre (L.) R. Br.	LECA1	Field Peppergrass
BRASSICACEAE	Lepidium densiflorum Schrad.	LEDE1	Peppergrass
BRASSICACEAE	Lesquerella montana (A. Gray) Wats.	LEMO1	Bladderpod
BRASSICACEAE	Nasturtium officinale R. Br.	NAOF1	Watercress
BRASSICACEAE	Physaria vitulifera Rydb.	PHVI1	Double Bladder-pod
BRASSICACEAE	Rorippa palustris (L.) Bess. ssp. hispida (Desv.) Jonsell	ROPA1	Bog Yellow Cress
BRASSICACEAE	Sisymbrium altissimum L.	SIAL1	Tumbling Mustard
BRASSICACEAE	Thlaspi arvense L.	THAR1	Field Penny Cress
CACTACEAE	Coryphantha missouriensis (Sweet) Britt. & Rose	COMI1	Nipple Cactus
CACTACEAE	Echinocereus viridiflorus Engelm.	ECVI1	Hedgehog Cactus
CACTACEAE	Opuntia fragilis (Nutt.) Haw.	OPFR1	Little Prickly Pear
CACTACEAE	Opuntia macrorhiza Engelm.	OPMA1	Twistspine Prickly Pear
CACTACEAE	Opuntia polyacantha Haw.	OPPO1	Plains Prickly Pear
CACTACEAE	Pediocactus simpsonii (Engelm.) Britt. & Rose	PESI1	Nipple Cactus
CALLITRICHACEAE	Callitricha verna L.	CAVE1	Water Starwort
CAMPANULACEAE	Campanula rotundifolia L.	CARO1	Harebell
CAMPANULACEAE	Lobelia siphilitica L. var. ludoviciana A. DC.	LOS1	Great Lobelia
CAMPANULACEAE	Triodanis leptocarpa (Nutt.) Nieuw.	TRLE1	Venus' Looking Glass
CANNABACEAE	Humulus lupulus L. var. lupuloides E. Small	HULU1	Common Hops
CAPPERACEAE	Polansia dodecandra (L.) DC. ssp. trachysperma (T. & G.) Iltis	PODO2	Clammy-weed
CAPRIFOLIACEAE	Symphoricarpos occidentalis Hook.	SYOC1	Western Snowberry
CAPRIFOLIACEAE	Symphoricarpos oreophilus Gray	SYOR1	Snowberry
CAPRIFOLIACEAE	Viburnum opulus L. var. americanum Ait	VIOP1	Highbush Cranberry
CARYOPHYLLACEAE	Arenaria fendleri A. Gray	ARFE2	Fendler's Sandwort
CARYOPHYLLACEAE	Cerastium arvense L.	CEAR1	Prairie Chickweed
CARYOPHYLLACEAE	Cerastium brachypodium (Engelm. ex A. Gray) Robins.	CEBR1	Short-stalked Chickweed
CARYOPHYLLACEAE	Cerastium vulgatum L.	CEVU1	Common Mouse-Ear
CARYOPHYLLACEAE	Conosilene conica (L.) Fourreau ssp. conoidea (L.) Love & Kjellqvist	COCO1	Community Campion
CARYOPHYLLACEAE	Paronychia jamesii T. & G.	PAJA1	James' Nailwort
CARYOPHYLLACEAE	Saponaria officinalis L.	SAOF1	Bouncing Bet
CARYOPHYLLACEAE	Silene antirrhina L.	SIAN1	Sleepy Catchfly
CARYOPHYLLACEAE	Silene drummondii Hook.	SIDR1	Campion
CARYOPHYLLACEAE	Silene pratensis (Raf.) Godr. & Gren	SIPR1	White Campion
CARYOPHYLLACEAE	Spergularia rubra (L.) K. Presl.	SPRU1	Sand Spurry
CARYOPHYLLACEAE	Stellaria longifolia Muhl. ex Willd.	STLO1	Long-leaved Stitchwort
CARYOPHYLLACEAE	Vaccaria pyramidata Medic.	VAPY1	Cow Cockle
CERATOPHYLLACEAE	Ceratophyllum demersum L.	CEDE1	Coontail

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CHENOPodiaceae	<i>Atriplex canescens</i> (Pursh.) Nutt.	ATCA1	Four-winged Saltbush
CHENOPodiaceae	<i>Chenopodium album</i> L.	CHAL1	Lamb's Quarters
CHENOPodiaceae	<i>Chenopodium atrovirens</i> Nutt.	CHAT1	Dark Goosefoot
CHENOPodiaceae	<i>Chenopodium berlandieri</i> Moq.	CHBE1	Pitseed Goosefoot
CHENOPodiaceae	<i>Chenopodium botrys</i> L.	CHBO1	Jerusalem Oak
CHENOPodiaceae	<i>Chenopodium dessicatum</i> A. Nels.	CHDE1	Desert goosefoot
CHENOPodiaceae	<i>Chenopodium fremontii</i> S. Wats.	CHFR1	Fremont Goosefoot
CHENOPodiaceae	<i>Chenopodium leptophyllum</i> Nutt. ex Moq.	CHLE2	Goosefoot
CHENOPodiaceae	<i>Chenopodium overi</i> Aellen	CHOV1	Overi's Goosefoot
CHENOPodiaceae	<i>Kochia scoparia</i> (L.) Schrad.	KOSC1	Kochia
CHENOPodiaceae	<i>Salsola iberica</i> Senn. & Pau.	SAIB1	Russian-Thistle
CLUSIACEAE	<i>Hypericum majus</i> (A. Gray) Britt.	HYMA1	Greater St. John's-wort
CLUSIACEAE	<i>Hypericum perforatum</i> L.	HYPE1	Common St. John's-wort
COMMELINACEAE	<i>Tradescantia occidentalis</i> (Britt.) Smyth	TROC1	Spiderwort
CONVOLVULACEAE	<i>Calystegia macouni</i> (Greene) Brummitt	CAMA1	Hedge Bindweed
CONVOLVULACEAE	<i>Convolvulus arvensis</i> L.	COAR1	Field Bindweed
CONVOLVULACEAE	<i>Evolvulus nuttallianus</i> R. & S.	EVNU1	Evolvulus
CRASSULACEAE	<i>Sedum lanceolatum</i> Torr.	SELA1	Stonecrop
CUPRESSACEAE	<i>Juniperus communis</i> L.	JUCO1	Common Juniper
CUPRESSACEAE	<i>Juniperus scopulorum</i> Sarg.	JUSC1	Rocky Mountain Juniper
CUSCUTACEAE	<i>Cuscuta approximata</i> Bab.	CUAP1	Dodder
CYPERACEAE	<i>Carex athrostachya</i> Olney	CAAT1	Sedge
CYPERACEAE	<i>Carex aurea</i> Nutt.	CAAU1	Sedge
CYPERACEAE	<i>Carex bebbii</i> (Bailey) Fern	CABE1	Sedge
CYPERACEAE	<i>Carex brevior</i> (Dew.) Mack. ex Lunell.	CABR1	Sedge
CYPERACEAE	<i>Carex douglasii</i> F. Boott.	CADO1	Sedge
CYPERACEAE	<i>Carex eleocharis</i> Bailey	CAEL1	Sedge
CYPERACEAE	<i>Carex emoryi</i> Dew.	CAEM1	Sedge
CYPERACEAE	<i>Carex filifolia</i> Nutt.	CAFI1	Sedge
CYPERACEAE	<i>Carex heliophila</i> Mack.	CAHE1	Sedge
CYPERACEAE	<i>Carex hystericina</i> Muhl. ex Willd.	CAHY1	Sedge
CYPERACEAE	<i>Carex interior</i> Bailey	CAIN1	Sedge
CYPERACEAE	<i>Carex lanuginosa</i> Michx.	CALA1	Sedge
CYPERACEAE	<i>Carex nebrascensis</i> Dew.	CANE1	Sedge
CYPERACEAE	<i>Carex oreocharis</i> Holm.	CAOR1	Sedge
CYPERACEAE	<i>Carex praegracilis</i> W. Boott.	CAPR1	Sedge
CYPERACEAE	<i>Carex rostrata</i> Stokes ex Willd.	CARO2	Sedge
CYPERACEAE	<i>Carex scoparia</i> Schkuhr. ex Willd.	CASC1	Sedge
CYPERACEAE	<i>Carex simulata</i> Mack.	CASI1	Sedge
CYPERACEAE	<i>Carex stipata</i> Muhl.	CAST1	Sedge
CYPERACEAE	<i>Carex vulpinoides</i> Michx.	CAVU1	Fox Sedge
CYPERACEAE	<i>Eleocharis acicularis</i> (L.) R. & S.	ELAC1	Spikerush
CYPERACEAE	<i>Eleocharis compressa</i> Sulliv.	ELCO1	Spikerush
CYPERACEAE	<i>Eleocharis macrostachya</i> Britt.	ELMA1	Spikerush
CYPERACEAE	<i>Eleocharis parvula</i> Link ex Boff. & Fingerbr. var. <i>anachaeta</i> (Torr.) Svens.	ELPA1	Spikerush
CYPERACEAE	<i>Scirpus acutus</i> Muhl.	SCAC1	Bulrush
CYPERACEAE	<i>Scirpus pallidus</i> (Britt.) Fern	SCPA1	Bulrush
CYPERACEAE	<i>Scirpus pungens</i> Vahl	SCPU1	Pungent Bulrush
CYPERACEAE	<i>Scirpus validus</i> Vahl.	SCVA1	Bulrush
ELAEAGNACEAE	<i>Elaeagnus angustifolia</i> L.	ELAN1	Russian Olive
EQUISETACEAE	<i>Equisetum arvense</i> L.	EQAR1	Field Horsetail
EQUISETACEAE	<i>Equisetum laevigatum</i> A. Br.	EQLA1	Smooth Horsetail
EQUISETACEAE	<i>Equisetum variegatum</i> Schleich.	EQVA1	Variegated Scouring Rush
EUPHORBIACEAE	<i>Euphorbia dentata</i> Michx.	EUDE1	Toothed Spurge
EUPHORBIACEAE	<i>Euphorbia fendleri</i> T. & G.	EUFE1	Fendler's Euphorbia
EUPHORBIACEAE	<i>Euphorbia marginata</i> Pursh.	EUMA1	Snow-on-the-Mountain
EUPHORBIACEAE	<i>Euphorbia robusta</i> (Engelm.) Small	EURO1	Spurge
EUPHORBIACEAE	<i>Euphorbia serpyllifolia</i> Pers.	EUSE1	Thyme-leaved Spurge
EUPHORBIACEAE	<i>Euphorbia spathulata</i> Lam.	EUSP1	Spurge
EUPHORBIACEAE	<i>Tragia ramosa</i> Nutt.	TRRA1	Noseburn
FABACEAE	<i>Amorpha fruticosa</i> L.	AMFR1	False Indigo
FABACEAE	<i>Amorpha nana</i> Nutt.	AMNA1	Dwarf Wild Indigo
FABACEAE	<i>Astragalus adsurgens</i> Pall. var. <i>robustior</i> Hook.	ASAD1	Standing Milkvetch
FABACEAE	<i>Astragalus agrestis</i> Dougl. ex G. Don	ASAG1	Field Milkvetch
FABACEAE	<i>Astragalus bisulcatus</i> (Hook.) A. Gray	ASBI1	Two-grooved Vetch

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FABACEAE	Astragalus canadensis L.	ASCA1	Canada Milk-vetch
FABACEAE	Astragalus crassicarpus Nutt.	ASCR1	Ground-plum
FABACEAE	Astragalus drummondii Dougl. ex Hook.	ASDR1	Drummond Milkvetch
FABACEAE	Astragalus flexuosus (Hook.) G. Don	ASFL1	Plaint Milkvetch
FABACEAE	Astragalus parryi Gray	ASPA1	Parry's Milkvetch
FABACEAE	Astragalus shortianus Nutt. ex T.&G.	ASSH1	Short's Milkvetch
FABACEAE	Astragalus spathulatus Sheld.	ASSP2	Draba Milk-Vetch
FABACEAE	Astragalus tridactylus Gray	ASTR1	Foothill Milkvetch
FABACEAE	Dalea candida Michx. ex Willd. var. oligophylla (Torr.) Shinners.	DACA1	White Prairie Clover
FABACEAE	Dalea purpurea Vent	DAPU1	Purple Prairie Clover
FABACEAE	Glycyrrhiza lepidota Pursh.	GLLE1	Wild Licorice
FABACEAE	Lathyrus eucosmus Butters and St. John	LAEU1	Purple Peavine
FABACEAE	Lotus corniculatus L.	LOC01	Birdfoot Trefoil
FABACEAE	Lupinus argenteus Pursh ssp. ingratus (Greene) Harmon	LUAR2	Silvery Lupine
FABACEAE	Lupinus argenteus Pursh var. argenteus	LUAR1	Black Medick
FABACEAE	Medicago lupulina L.	MELU1	Alfalfa
FABACEAE	Medicago sativa L. ssp. sativa	MESA1	
FABACEAE	Melilotus alba Medic.	MEAL1	White Sweetclover
FABACEAE	Melilotus officinalis (L.) Pall.	MEOF1	Yellow Sweetclover
FABACEAE	Oxytropis lambertii Pursh.	OXLA1	Purple Locoweed
FABACEAE	Psoralea tenuiflora Pursh.	PSTE1	Wild Alfalfa
FABACEAE	Robinia pseudo-acacia L.	ROPS1	Black Locust
FABACEAE	Thermopsis rhombifolia var. divaricarpa (Nels.) Isely	THRH1	Golden Banner
FABACEAE	Trifolium hybridum L.	TRHY1	Alsike Clover
FABACEAE	Trifolium pratense L.	TRPR1	Red Clover
FABACEAE	Vicia americana Muhl. ex Willd.	VIAM1	American Vetch
FUMARIACEAE	Fumaria vaillantii Lois	FUVA1	Fumitory
GENTIANACEAE	Gentiana affinis Griseb.	GEAF1	Northern Gentian
GENTIANACEAE	Swertia radiata (Kell.) O. Ktze.	SWRA1	Green Gentian
GERANIACEAE	Erodium cicutarium (L.) L'Her.	ERCI1	Filaria
GERANIACEAE	Geranium caespitosum James ssp. caespitosum	GECA1	Common Wild Geranium
GROSSULARIACEAE	Ribes aureum Pursh	RIAU1	Golden Currant
GROSSULARIACEAE	Ribes cereum Dougl.	RICE1	Western Red Currant
HALORAGACEAE	Myriophyllum exalbescens Fern.	MYEX1	American Milfoil
HYDROPHYLACEAE	Hydrophyllum fendleri (Gray) Heller	HYFE1	Waterleaf
HYDROPHYLACEAE	Phacelia heterophylla Pursh.	PHHE1	Scorpionweed
IRIDACEAE	Iris missouriensis Nutt.	IRMI1	Western Blue Flag
IRIDACEAE	Sisyrinchium montanum Greene	SIMO1	Blue-eyed Grass
JUNCACEAE	Juncus articulatus L.	JUAR1	Articulate Rush
JUNCACEAE	Juncus balticus Willd.	JUBA1	Baltic Rush
JUNCACEAE	Juncus bufonius L.	JUBU1	Toad Rush
JUNCACEAE	Juncus dudleyi Wieg.	JUDU1	Dudley Rush
JUNCACEAE	Juncus ensifolius Wikst. var. montanus (Engelm.) C. L. Hitchc.	JUEN1	Rush
JUNCACEAE	Juncus interior Wieg.	JUIN1	Inland Rush
JUNCACEAE	Juncus longistylis Torr.	JUL01	Rush
JUNCACEAE	Juncus nodosus L.	JUNO1	Knotted Rush
JUNCACEAE	Juncus torreyi Cov.	JUTO1	Torrey's Rush
JUNCACEAE	Juncus tracyi Rydb.	JUTR1	Tracy Rush
LAMIACEAE	Dracocephalum parviflorum Nutt.	DRPA1	Dragonhead
LAMIACEAE	Hedeoma hispidum Pursh.	HEHI1	Rough False Pennyroyal
LAMIACEAE	Lycopus americanus Muhl. ex Barton	LYAM1	American Bugleweed
LAMIACEAE	Marrubium vulgare L.	MAVU1	Common Horehound
LAMIACEAE	Mentha arvensis L.	MEAR1	Field Mint
LAMIACEAE	Monarda fistulosa L. var. menthifolia (Grah.) Fern.	MOF11	Wild Bergamot
LAMIACEAE	Monarda pectinata Nutt.	MOPE1	Spotted Bee-Balm
LAMIACEAE	Nepeta cataria L.	NECA1	Catnip
LAMIACEAE	Prunella vulgaris L.	PRVU1	Selfheal
LAMIACEAE	Salvia reflexa Hornem.	SARE1	Lance-leaved Sage
LAMIACEAE	Scutellaria brittonii Porter	SCBR1	Britton's Skullcap
LAMIACEAE	Stachys palustris L. ssp. pilosa (Nutt.) Epling	STPA2	Hedge Nettle
LEMNACEAE	Lemna minor L.	LEMI1	Duckweed
LILIACEAE	Allium cernuum Roth	ALCE1	Wild Onion
LILIACEAE	Allium geyeri S. Wats.	ALGE1	Geyer's Onion
LILIACEAE	Allium textile A. Nels. & Macbr.	ALTE1	Wild White Onion
LILIACEAE	Asparagus officinalis L.	ASOF1	Asparagus

Plant Species Known to Occur at Rocky Flats Environmental Technology Site, May 1999 (cont.)

Family	Scientific Name	Speccode	Common Name
LILIACEAE	Calochortus gunnisonii S. Wats.	CAGU1	Sego Lily
LILIACEAE	Leucocrinum montanum Nutt.	LEMO2	Mountain Lily
LILIACEAE	Smilacina stellata (L.) Desf.	SMST1	Spikenard
LILIACEAE	Zigadenus venenosus Wats. var. gramineus (Rydb.) Walsh ex Peck	ZIVE1	Death Camass
LINACEAE	Linum perenne L. var. lewisii (Pursh.) Eat. & Wright	LIPE1	Blue Flax
LINACEAE	Linum pratense (Nort.) Small	LIPR1	Norton's Flax
LYTHRACEAE	Ammania robusta Herr & Regel.	AMRO1	Robust Toothcup
LYTHRACEAE	Lythrum alatum Pursh.	LYAL1	Winged Loosestrife
MALVACEAE	Malva neglecta Wallr.	MANE1	Common Mallow
MALVACEAE	Sidalcea candida Gray	SICA1	White Checkermallow
MALVACEAE	Sidalcea neomexicana Gray	SINE1	New Mexico Checkmallow
MALVACEAE	Sphaeralcea coccinea (Pursh.) Rydb.	SPCO1	Red False Mallow
NYCTAGINACEAE	Mirabilis hirsuta (Pursh.) MacM.	MIH1	Hairy Four-O'Clock
NYCTAGINACEAE	Mirabilis linearis (Pursh.) Heimerl	MILI1	Narrowleaf Four O'Clock
NYCTAGINACEAE	Mirabilis nyctaginea (Michx.) MacM.	MINY1	Wild Four-O'Clock
ONAGRACEAE	Calylophus serrulatus (Nutt.) Raven	CASE2	Plains Yellow Primrose
ONAGRACEAE	Epilobium ciliatum Raf. ssp. glandulosum (Lehm.) Hock & Raven	EPCI1	Willow Herb
ONAGRACEAE	Epilobium paniculatum Nutt.	EPPA1	Willow Herb
ONAGRACEAE	Gaura coccinea Pursh.	GACO1	Scarlet Gaura
ONAGRACEAE	Gaura parviflora Dougl.	GAPA1	Velvety Gaura
ONAGRACEAE	Oenothera flava (A. Nels.) Garrett	OEFL1	Evening Primrose
ONAGRACEAE	Oenothera howardii (A. Nels.) W. L. Wagner	OEH01	Yellow Stemless Evening Pr
ONAGRACEAE	Oenothera villosa Thunb. ssp. strigosa (Rydb.) Dietrich & Raven	OEV1	Common Evening Primrose
ORCHIDACEAE	Habenaria hyperborea (L.) R. Br.	HAHY1	Northern Green Orchid
OROBANCHACEAE	Orobanche fasciculata Nutt.	ORFA1	Broomrape
OXALIDACEAE	Oxalis dillenii Jacq.	OXDI1	Gray-Green Wood Sorrel
PAPAVERACEAE	Argemone polyanthemos (Fedde) G. Ownbey	ARPO1	Prickly Poppy
PINACEAE	Picea pungens Engelm.	PIPU1	Blue Spruce
PINACEAE	Pinus ponderosa Laws	PIPO1	Ponderosa Pine
PINACEAE	Pseudotsuga menziesii (Mirb.) Franco	PSME1	Douglas-Fir
PLANTAGINACE	Plantago lanceolata L.	PLLA1	English Plantain
PLANTAGINACE	Plantago major L.	PLMA1	Common Plantain
PLANTAGINACE	Plantago patagonica Jacq.	PLPA1	Patagonian Plantain
POACEAE	Aegilops cylindrica Host	AECY1	Jointed Goatgrass
POACEAE	Agropyron caninum (L.) Beauv. ssp. majus (Vasey) C. L. Hitchc.	AGCA1	Slender Wheatgrass
POACEAE	Agropyron cristatum (L.) Gaertn.	AGCR1	Crested Wheatgrass
POACEAE	Agropyron dasystachyum (Hook.) Scribn.	AGDA1	
POACEAE	Agropyron desertorum (Fisch.) Schult.	AGDE1	Crested Wheatgrass
POACEAE	Agropyron elongatum (Host) Beauv.	AGE1	Tall Wheatgrass
POACEAE	Agropyron griffithsii Scribn. & Smith	AGGR1	
POACEAE	Agropyron intermedium (Host) Beauv.	AGIN1	Intermediate Wheatgrass
POACEAE	Agropyron repens (L.) Beauv.	AGRE1	Quackgrass
POACEAE	Agropyron smithii Rydb.	AGSM1	Western Wheatgrass
POACEAE	Agropyron spicatum (Pursh) Scribn. and Sm.	AGSP1	Bluebunch Wheatgrass
POACEAE	Agrostis scabra Willd.	AGSC1	Ticklegrass
POACEAE	Agrostis stolonifera L.	AGST1	Redtop
POACEAE	Alopecurus geniculatus L.	ALGE2	Marsh Foxtail
POACEAE	Andropogon gerardii Vitman	ANGE1	Big Bluestem
POACEAE	Andropogon scoparius Michx.	ANSC1	Little Bluestem
POACEAE	Apera interrupta (L.) Beauvois	APIN1	Italian Windgrass
POACEAE	Aristida basiramea Engelm. ex Vasey var. basiramea	ARBA1	Forktip Threeawn
POACEAE	Aristida purpurea Nutt. var. longiseta (Steud.) Vasey	ARFE1	Fendler Threeawn
POACEAE	Aristida purpurea Nutt. var. robusta (Merrill) A. Holmgren & N. Holmgr	ARLO1	Red Threeawn
POACEAE	Avena fatua var. sativa (L.) Hausskn.	AVFA1	Cultivated Oats
POACEAE	Bouteloua curtipendula (Michx.) Torr.	BOCU1	Side-oats Gram
POACEAE	Bouteloua gracilis (H. B. K.) Lag ex Griffiths	BOGR1	Blue Grama
POACEAE	Bouteloua hirsuta Lag	BOHI1	Hairy Grama
POACEAE	Bromus briziformis F. & M.	BRBR1	Rattlesnake Grass
POACEAE	Bromus inermis Leyss. ssp. inermis	BRIN1	Smooth Brome
POACEAE	Bromus japonicus Thunb. ex Murr.	BRJA1	Japanese Brome
POACEAE	Bromus tectorum L.	BRTE1	Downy Brome
POACEAE	Buchloe dactyloides (Nutt.) Engelm.	BUDA1	Buffalo-grass
POACEAE	Calamagrostis stricta (Timm.) Koel	CAST2	Northern Reedgrass
POACEAE	Cenchrus longispinus (Hack.) Fern	CELO1	Field Sandbur
POACEAE	Ceratochloa marginata (Nees ex Stued.) Jackson	CEMA1	Rescuegrass

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Family	Scientific Name	Speccode	Common Name
POACEAE	Dactylis glomerata L.	DAGL1	Orchardgrass
POACEAE	Danthonia spicata (L.) Beauv. ex R. & S.	DASP1	Poverty Oatgrass
POACEAE	Dichanthelium linearifolium (Scribn.) Gould	DILI1	Slimleaf Dichanthelium
POACEAE	Dichanthelium oligosanthes (Schultz) Gould var. scribnerianum (Nash) G	DIOL1	Scribner Dichanthelium
POACEAE	Digitaria sanguinalis (L.) Scop.	DISA1	Hairy Crabgrass
POACEAE	Echinochloa crusgalli (L.) Beauv.	ECCR1	Barnyard Grass
POACEAE	Elymus canadensis L.	ELCA1	Canada Wild Rye
POACEAE	Elymus juncea Fisch.	ELJU1	Russian Wild Rye
POACEAE	Eragrostis cilianensis (All.) E. Mosher	ERCI2	Stinkgrass
POACEAE	Eragrostis curvula (Schrad.) Nees	ERCU1	Weeping Lovegrass
POACEAE	Eragrostis pilosa (L.) Beauv.	ERP1	India Lovegrass
POACEAE	Festuca octoflora Walt.	FEOC1	Six-weeks Fescue
POACEAE	Festuca ovina L. var. rydbergii St. Yves	FEOV1	Sheep's Fescue
POACEAE	Festuca pratensis Huds.	FEPR1	Meadow Fescue
POACEAE	Glyceria grandis S. Wats. ex A. Gray	GLGR1	Tall Mannagrass
POACEAE	Glyceria striata (Lam.) Hitchc.	GLST1	Fowl Mannagrass
POACEAE	Hordeum brachyantherum Nevski	HOBR1	Meadow Barley
POACEAE	Hordeum jubatum L.	HOJU1	Foxtail Barley
POACEAE	Koeleria pyramidata (Lam.) Beauv.	KOPY1	Junegrass
POACEAE	Leersia oryzoides (L.) Sw.	LEOR1	Rice Cutgrass
POACEAE	Lolium perenne L.	LOPE1	Ryegrass
POACEAE	Muhlenbergia asperifolia (Nees. & Mey.) Parodi	MUAS1	Scratchgrass
POACEAE	Muhlenbergia filiformis (Thurb.) Rydb.	MUF11	Muhly
POACEAE	Muhlenbergia montana (Nutt.) Hitchc.	MUMO1	Mountain Muhly
POACEAE	Muhlenbergia racemosa (Michx.) B. S. P.	MURA1	Marsh Muhly
POACEAE	Muhlenbergia wrightii Vasey	MUWR1	Spike Muhly
POACEAE	Oryzopsis hymenoides (R. & S.) Ricker	ORHY1	Indian Ricegrass
POACEAE	Panicum capillare L.	PACA1	Witchgrass
POACEAE	Panicum virgatum L.	PAVI1	Switchgrass
POACEAE	Phalaris arundinacea L.	PHAR1	Reed Canarygrass
POACEAE	Phleum pratense L.	PHPR1	Timothy
POACEAE	Phragmites australis (Cav.) Trin. ex Steud.	PHAU1	Common Reed
POACEAE	Poa bulbosa L.	POBU1	Bulbous Bluegrass
POACEAE	Poa canbyi (Scribn.) Piper	POCA1	Canby's Bluegrass
POACEAE	Poa compressa L.	POCO1	Canada Bluegrass
POACEAE	Poa fendleriana (Steud.) Vasey	POFE1	Mutongrass
POACEAE	Poa palustris L.	POPA1	Fowl Bluegrass
POACEAE	Poa pratensis L.	POPR1	Kentucky Bluegrass
POACEAE	Polypogon monspeliensis (L.) Desf.	POMO1	Rabbitfoot Grass
POACEAE	Schedonnardus paniculatus (Nutt.) Trel.	SCPA2	Tumblegrass
POACEAE	Secale cereale L.	SECE1	Rye
POACEAE	Setaria viridis (L.) Beauv.	SEVI1	Green Foxtail
POACEAE	Sitanion hystrix (Nutt.) Sm. var. brevifolium (Sm.) Hitchc.	SIHY1	Squirreltail
POACEAE	Sorghastrum nutans (L.) Nash	SONU1	Indian-grass
POACEAE	Spartina pectinata Link	SPPE1	Prairie Cordgrass
POACEAE	Sphenopholis obtusata (Michx.) Scribn.	SPOB1	Prairie Wedgegrass
POACEAE	Sporobolus asper (Michx.) Kunth	SPAS1	Rough Dropseed
POACEAE	Sporobolus cryptandrus (Torr.) A. Gray	SPCR1	Sand Dropseed
POACEAE	Sporobolus heterolepis (A. Gray) A. Gray	SPHE1	Prairie Dropseed
POACEAE	Sporobolus neglectus Nash	SPNE1	Poverty Grass
POACEAE	Stipa comata Trin. & Rupr.	STCO1	Needle-and-thread
POACEAE	Stipa neomexicana (Thur.) Scribn.	STNE1	New Mexico Feather Grass
POACEAE	Stipa spartea Trinius	STSP1	Porcupine-grass
POACEAE	Stipa viridula Trin.	STVI1	Green Needlegrass
POACEAE	Triticum aestivum L.	TRAЕ1	Wheat
POACEAE	X Agrohordeum macounii (Vasey) Lepage	AGMA1	
POLEMONIACEAE	Collomia linearis Nutt.	COLI1	Collomia
POLEMONIACEAE	Gilia opthalmodoides Brand. ssp. clokeyi (Mason) A. & V. Grant	GIOP1	Gilia
POLEMONIACEAE	Ipomopsis spicata (Nutt.) V. Grant ssp. spicata	IPSP1	Spike Gilia
POLEMONIACEAE	Microsteris gracilis (Hook.) Greene	MIGR1	
POLEMONIACEAE	Navarretia minima Nutt.	NAMI1	Navarretia
POLYGONACEAE	Eriogonum alatum Torr.	ERAL1	Winged Eriogonum
POLYGONACEAE	Eriogonum effusum Nutt.	EREF1	Spreading Wild Buckwheat
POLYGONACEAE	Eriogonum jamesii Benth.	ERJA1	James' Wild Buckwheat
POLYGONACEAE	Eriogonum umbellatum Torr.	ERUM1	Sulphur Flower

Plant Species Known to Occur at Rocky Flats Environmental Technology Site, May 1999 (cont.)

Family	Scientific Name	Speccode	Common Name
POLYGONACEAE	Polygonum arenastrum Jord. ex Bor.	POAR1	Knotweed
POLYGONACEAE	Polygonum convolvulus L.C394	POCO2	Wild Buckwheat
POLYGONACEAE	Polygonum douglasii Greene	PODO1	Knotweed
POLYGONACEAE	Polygonum hydropiper L.	POHY1	Water Pepper
POLYGONACEAE	Polygonum lapathifolium L.	POLA1	Pale Smartweed
POLYGONACEAE	Polygonum pensylvanicum L.	POPE1	Pennsylvania Smartweed
POLYGONACEAE	Polygonum persicaria L.	POPE2	Lady's Thumb
POLYGONACEAE	Polygonum ramosissimum Michx.	PORA1	Knotweed
POLYGONACEAE	Polygonum sawatchense Small	POSA1	Knotweed
POLYGONACEAE	Rumex acetosella L.	RUAC1	Sheep Sorrel
POLYGONACEAE	Rumex crispus L.	RUCR1	Curly Dock
POLYGONACEAE	Rumex maritimus L.	RUMA1	Golden Dock
POLYGONACEAE	Rumex obtusifolius L.	RUOB1	Bitter Dock
POLYGONACEAE	Rumex salicifolius Weinm. ssp. triangulivalvis Danser	RUSA1	Willow Dock
POLYPODIACEAE	Cystopteris fragilis (L.) Bernh.	CYFR1	Fragile Fern
PORTULACACEAE	Claytonia rosea Rydb.	CLRO1	Spring Beauty
PORTULACACEAE	Portulaca oleracea L.	POOL1	Common Purslane
PORTULACACEAE	Talinum parviflorum Nutt.	TAPA1	Prairie Fameflower
POTAMOGETONACEAE	Potamogeton foliosus Raf.	POFO1	Leafy Pondweed
POTAMOGETONACEAE	Potamogeton natans L.	PONA1	Floatingleaf Pondweed
PRIMULACEAE	Androsace occidentalis Pursh.	ANOC1	Western Rock Jasmine
PRIMULACEAE	Dodecatheon pulchellum (Raf.) Merrill	DOPU1	Shooting Star
PRIMULACEAE	Lysimachia ciliata L.	LYCI1	Fringed Lostrife
RANUNCULACEAE	Anemone cylindrica A. Gray	ANCY1	Candle Anemone
RANUNCULACEAE	Anemone patens L.	ANPA2	Pasque-flower
RANUNCULACEAE	Clematis hirsutissima Pursh	CLHI1	Hairy Clematis
RANUNCULACEAE	Clematis ligusticifolia Nutt.	CLLI1	Western Clematis
RANUNCULACEAE	Delphinium nuttalianum Pritz. ex Walpers	DENU1	Blue Larkspur
RANUNCULACEAE	Delphinium virescens Nutt. ssp. penardii (Huth) Ewan	DEV11	Prairie Larkspur
RANUNCULACEAE	Myosurus minimus L.	MYMI1	Mousetail
RANUNCULACEAE	Ranunculus macounii Britt.	RAMA1	Macoun's Buttercup
RANUNCULACEAE	Ranunculus scleratus L.	RASC1	Cursed Crowfoot
RANUNCULACEAE	Ranunculus trichophyllus Chaix	RATR1	Hairy Leaf Buttercup
RANUNCULACEAE	Thalictrum dasycarpum Fisch. & Ave-Lall	THDA1	Purple Meadow Rue
RHAMNACEAE	Ceanothus herbaceus Raf. var. pubescens (T. & G.)	CEHE1	New Jersey Tea
ROSACEAE	Agrimonia striata Michx.	AGST2	Striate Agrimony
ROSACEAE	Amelanchier alnifolia Nutt.	AMAL1	Saskatoon Service-berry
ROSACEAE	Crataegus erythropoda Ashe	CRER1	Hawthorne
ROSACEAE	Crataegus succulenta Link var. occidentalis (Britton) E. J. Palm.	CRSU1	Hawthorn
ROSACEAE	Geum aleppicum Jacq.	GEAL1	Yellow Avens
ROSACEAE	Geum macrophyllum Willd.	GEMA1	Large-leaved Avens
ROSACEAE	Physocarpus monogynus (Torr.) Coul.	PHMO1	Mountain Ninebark
ROSACEAE	Physocarpus opulifolius (L.) Raf.	PHOP1	Ninebark
ROSACEAE	Potentilla arguta Pursh	POAR2	Tall Cinquefoil
ROSACEAE	Potentilla fissa Nutt.	POF11	Cinquefoil
ROSACEAE	Potentilla gracilis Dougl. ex Hook. var. glabrata (Lehm.) C. L. Hitchc.	POGR1	Cinquefoil
ROSACEAE	Potentilla hippiana Lehm.	POHI1	Wooly Cinquefoil
ROSACEAE	Potentilla norvegica L.	PONO1	Norwegian Cinquefoil
ROSACEAE	Potentilla paradoxa Nutt.	POPA2	Bushy Cinquefoil
ROSACEAE	Potentilla pensylvanica L.	POPE4	Cinquefoil
ROSACEAE	Potentilla pulcherrima x hippiana	POP11	Hybrid Cinquefoil
ROSACEAE	Potentilla rivalis Nutt.	PORI1	Cinquefoil
ROSACEAE	Prunus americana Marsh.	PRAM1	Wild Plum
ROSACEAE	Prunus pumila L. var. besseyi (Bailey) Gl.	PRPU1	Sand Cherry
ROSACEAE	Prunus virginiana L. var. melanocarpa (A. Nels.) Sarg.	PRV11	Chokecherry
ROSACEAE	Pyrus malus L.	PYMA1	Apple
ROSACEAE	Rosa acicularis Lindl.	ROAC1	Prickly Wild Rose
ROSACEAE	Rosa arkansana Porter	ROAR1	Prairie Wild Rose
ROSACEAE	Rosa woodsi Lindl.	ROWO1	Western Wild Rose
ROSACEAE	Rubus deliciosus Torr.	RUDE1	Boulder Raspberry
ROSACEAE	Rubus idaeus L. ssp. sachalinensis (Levl.) Focke var. sachalinensis	RUID1	Raspberry
ROSACEAE	Sanguisorba minor Scop.	SAMI1	Burnet
ROSACEAE	Sorbus scopulina Greene	SOSC1	Mountain Ash
RUBIACEAE	Galium aparine L.	GAAP1	Catchweed Bedstraw
RUBIACEAE	Galium septentrionale Roemer & Schultes	GASE1	Northern Bedstraw

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Family	Scientific Name	Speccode	Common Name
SALICACEAE	<i>Populus alba</i> L.	POAL1	Silver Poplar
SALICACEAE	<i>Populus angustifolia</i> James	POAN3	Narrow-leaved Cottonwood
SALICACEAE	<i>Populus deltoides</i> Marsh. ssp. <i>monilifera</i> (Ait.) Eckenw.	PODE1	Plains Cottonwood
SALICACEAE	<i>Populus x acuminata</i> Rydb.	POAC1	Lanceleaf Cottonwood
SALICACEAE	<i>Salix amygdaloides</i> Anderss.	SAAM1	Peach-leaf Willow
SALICACEAE	<i>Salix exigua</i> Nutt. ssp. <i>exigua</i>	SAEX2	Coyote Willow
SALICACEAE	<i>Salix exigua</i> Nutt. ssp. <i>interior</i> (Rowlee) Cronq.	SAEX1	Sandbar Willow
SALICACEAE	<i>Salix fragilis</i> L.	SAFR1	Crack Willow
SALICACEAE	<i>Salix irrorata</i> Andersson	SAIR1	
SALICACEAE	<i>Salix lutea</i> Nutt.	SALU1	Yellow Willow
SANTALACEAE	<i>Comandra umbellata</i> (L.) Nutt.	COUM1	Bastard Toadflax
SAXIFRAGACEAE	<i>Heuchera parvifolia</i> Nutt. ex T. & G.	HEPA1	Alumroot
SAXIFRAGACEAE	<i>Saxifraga rhomboidea</i> Greene	SARH1	Diamondleaf Saxifrage
SCROPHULARIACEAE	<i>Castilleja integra</i> A. Gray	CAIN2	Orange Paintbrush
SCROPHULARIACEAE	<i>Castilleja sessiliflora</i> Pursh.	CASE3	Downy Paintbrush
SCROPHULARIACEAE	<i>Collinsia parviflora</i> Doug. ex Lindl.	COPA1	Blue Lips
SCROPHULARIACEAE	<i>Gratiola neglecta</i> Torr.	GRNE1	Hedge Hyssop
SCROPHULARIACEAE	<i>Linaria dalmatica</i> (L.) Mill.	LIDA1	Toadflax
SCROPHULARIACEAE	<i>Linaria vulgaris</i> Hill	LIVU1	Butter-and-eggs
SCROPHULARIACEAE	<i>Mimulus floribundus</i> Dougl. ex Lindl.	MIFL1	Monkey Flower
SCROPHULARIACEAE	<i>Mimulus glabratus</i> H. B. K. var. <i>fremontii</i> (Benth.) A. L. Grant	MIGL1	Roundleaf Monkey-flower
SCROPHULARIACEAE	<i>Penstemon albidus</i> Nutt.	PEAL1	White Beardtongue
SCROPHULARIACEAE	<i>Penstemon secundiflorus</i> Benth.	PESE1	Penstemon
SCROPHULARIACEAE	<i>Penstemon strictus</i> Bentham in De Candolle	PEST1	Rocky Mountain Penstemon
SCROPHULARIACEAE	<i>Penstemon virens</i> Penn.	PEVI1	Slender Penstemon
SCROPHULARIACEAE	<i>Penstemon virgatus</i> Gray ssp. <i>asa-grayi</i> Crosswhite	PEVI2	Penstemon
SCROPHULARIACEAE	<i>Scrophularia lanceolata</i> Pursh.	SCLA2	Figwort
SCROPHULARIACEAE	<i>Verbascum blattaria</i> L.	VEBL1	Moth Mullein
SCROPHULARIACEAE	<i>Verbascum thapsus</i> L.	VETH1	Common Mullein
SCROPHULARIACEAE	<i>Veronica americana</i> (Raf.) Schwein. ex Benth.	VEAM1	Brooklime Speedwell
SCROPHULARIACEAE	<i>Veronica anagallis-aquatica</i> L.	VEAN1	Water Speedwell
SCROPHULARIACEAE	<i>Veronica catenata</i> Penn.	VECA1	Catenate Ironweed
SCROPHULARIACEAE	<i>Veronica peregrina</i> L. var. <i>xalapensis</i> (H. B. K.) St. John & Warren	VEPE1	Purslane Speedwell
SELAGINELLACEAE	<i>Selaginella densa</i> Rydb.	SEDE1	Spikemoss
SMILACACEAE	<i>Smilax herbacea</i> L. var. <i>lasioneura</i> (Small) Rydb..	SMHE1	Carrion Flower
SOLANACEAE	<i>Physalis heterophylla</i> Nees	PHHE2	Clammy Ground cherry
SOLANACEAE	<i>Physalis virginiana</i> P. Mill.	PHVI2	Virginia Ground Cherry
SOLANACEAE	<i>Quinula lobata</i> (Torr.) Raf.	QULO1	Purple Ground Cherry
SOLANACEAE	<i>Solanum rostratum</i> Dun.	SORO1	Buffalo Bur
SOLANACEAE	<i>Solanum triflorum</i> Nutt.	SOTR1	Cut-leaved Nightshade
TAMARICACEAE	<i>Tamarix ramosissima</i> Ledeb.	TARA1	Salt Cedar
TYPHACEAE	<i>Typha angustifolia</i> L.	TYAN1	Narrow-leaved Cattail
TYPHACEAE	<i>Typha latifolia</i> L.	TYLA1	Common Cattail
ULMACEAE	<i>Ulmus pumila</i> L.	ULPU1	Siberian Elm
URTICACEAE	<i>Parietaria pensylvanica</i> Muhl. ex Willd.	PAPE1	Pennsylvania Pellitory
URTICACEAE	<i>Urtica dioica</i> L. ssp. <i>gracilis</i> (Ait.) Seland.	URDI1	Stinging Nettle
VERBENACEAE	<i>Lippia cuneifolia</i> (Torr.) Steud.	LICU1	Fog-fruit
VERBENACEAE	<i>Verbena bracteata</i> Lag. & Rodr.	VEBR1	Prostrate Vervain
VERBENACEAE	<i>Verbena hastata</i> L.	VEHA1	Blue Vervain
VIOLACEAE	<i>Hybanthus verticillatus</i> (Ort.) Baill.	HYVE1	Nodding Green Violet
VIOLACEAE	<i>Viola nuttallii</i> Pursh.	VINU1	Yellow Prairie Violet
VIOLACEAE	<i>Viola rydbergii</i> Greene	VIRY1	Rydberg's Violet
VIOLACEAE	<i>Viola scopulorum</i> (Gray) Greene	VISC1	Colorado Violet
VIOLACEAE	<i>Viola sororia</i> Willd.	VISO1	Northern Bog Violet
VITACEAE	<i>Vitis riparia</i> Michx.	VIRI1	River-bank Grape
ZYGOPHYLLACEAE	<i>Tribulus terrestris</i> L.	TRTE1	Puncture Vine